



US007994393B2

(12) **United States Patent**
Lam et al.

(10) **Patent No.:** **US 7,994,393 B2**

(45) **Date of Patent:** **Aug. 9, 2011**

(54) **METHOD TO IMPROVE PLANT RESISTANCE TO INFECTIONS**

(75) Inventors: **Hon-Ming Lam**, Hong Kong (CN); **Sai Ming Samuel Sun**, Hong Kong (CN)

(73) Assignee: **The Chinese University of Hong Kong**, Hong Kong (CN)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 235 days.

(21) Appl. No.: **12/145,435**

(22) Filed: **Jun. 24, 2008**

(65) **Prior Publication Data**

US 2010/0313296 A1 Dec. 9, 2010

Related U.S. Application Data

(60) Provisional application No. 60/947,365, filed on Jun. 29, 2007, provisional application No. 60/947,590, filed on Jul. 2, 2007.

(51) **Int. Cl.**
A01H 5/00 (2006.01)
C12N 15/09 (2006.01)
C12N 15/82 (2006.01)

(52) **U.S. Cl. 800/279; 800/278; 800/298; 435/320.1; 435/468; 435/419; 435/418**

(58) **Field of Classification Search** None
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

2002/0031818 A1 3/2002 Ronai et al.
2002/0119547 A1 8/2002 Curtis et al.
2006/0123505 A1* 6/2006 Kikuchi et al. 800/278

OTHER PUBLICATIONS

International Search Report and Written Opinion for PCT/US08/68189, mailed Mar. 10, 2009, 7 pages.
Stone et al., Plant Physiology (2005) 137:13-30.

* cited by examiner

Primary Examiner — Medina A Ibrahim

(74) *Attorney, Agent, or Firm* — Morrison & Foerster LLP

(57) **ABSTRACT**

The disclosed invention relates to expression systems that effect production of a protein in plants that confers resistance to trauma. The expression systems are used to modify plants to improve their resistance to infections and wounding.

7 Claims, 12 Drawing Sheets

atgccagccccttcgcttctctcatggccgctcattgggctccttgccattcaattgttgca
M P A P S L P H G R H W A P C H S I V A
gcgccgttgcttattgcggttgagctgctgctttgcatatctctgaaaagttgagagtt
A P L L I A F E L L L C I Y L E S L R V
aaaagtaagccgactggtgattgaagattgtattccttctcttctggcctttgaagtg
K S K P T V D L K I V F L P L L A F E V
attattcttggttgacaatttcagaatgtgtagagctttaatgccaggagatgaagaaagt
I I L V D N F R M C R A L M P G D E E S
atgagcgatgaagctatttgggagacacttctcacttttgggttgcaatttctatggtg
M S D E A I W E T L P H F W V A I S M V
ttcttatagctgctacaaccttcacacttttgaagctgtctgggtgatgttgggtgctttg
F L I A A T T F T L L K L S G D V G A L
ggatgggtgggatttgtttataaattatggaatcgcgagtgtttgcatttcttgtttgt
G W W D L F I N Y G I A E C F A F L V C
actagatggtttaatcccagatgattcataaatctcctaactctggggaggctagctcatca
T R W F N P M I H K S P N P G E A S S S
tcagcggcaattagataaccgtgattgggagagtggtcttctctctccatcactagaagat
S A A I R Y R D W E S G L L L P S L E D
catgaacaagagaggctctgtggtcttctctgacataggcggtcacgtaatgaaaatacca
H E Q E R L C G L P D I G G H V M K I P
ctggtgattttccaagtttggcttggatgagccttggagggtacgcctcctagtgctcag
L V I F Q V L L C M R L E G T P P S A Q
tatattccgatatttgcactgttctctccactatttattttacaaggcgtgggtgctctt
Y I P I F A L F S P L F I L Q G A G V L
ttctctctagcaagattgttggagaaggttgttctactattacgaaatggaccagttagt
F S L A R L L E K V V L L L R N G P V S
cctaattaccttacaatctcatcaaaagtcogtgattgctttgcttttctcatcggtgt
P N Y L T I S S K V R D C F A F L H R G
tcaaggcttcttgggtgggtgctattgatgaaggcagcaagaagagcaagccccggtt
S R L L G W W S I D E G S K E E Q A R L
ttctatactgaatctactgggtacaacacatttgggttatccacctgaggtagtcagg
F Y T E S T G Y N T F C G Y P P E V V R
aaaatgcctaagagggatcttgcagaagaggtatggaggctccaagcagctttgggagag
K M P K R D L A E E V W R L Q A A L G E
caatcagaaattaccaaatgtaccaagcaggaatttgaaggcttcaaatgagaaggtt
Q S E I T K C T K Q E F E R L Q N E K V
cttgttaggatttgcctacgagggggagatatgcatggtcttacttcttggccggcacaga
L C R I C Y E G E I C M V L L P C R H R
acattatgcaagacttgttctgataagtgcaagaaatgtccaatctgccgtgtgccatt
T L C K T C S D K C K K C P I C R V P I
gaagaacgcctgcccgtatgatgtttaa
E E R M P V Y D V -

Figure 1

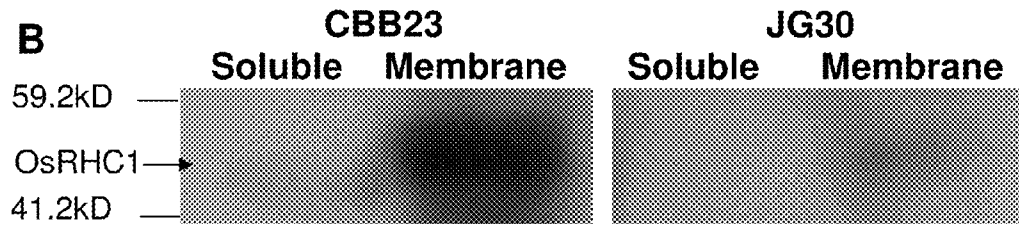


Figure 2B

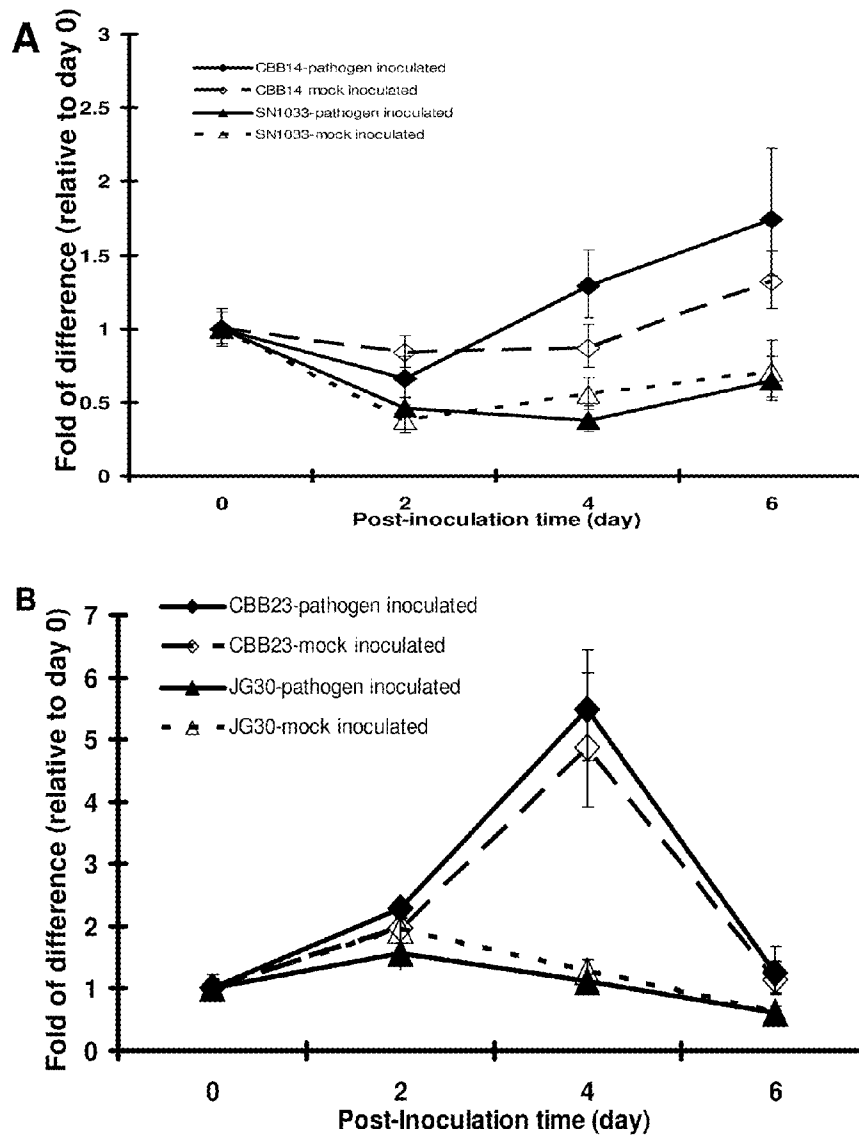


Figure 3

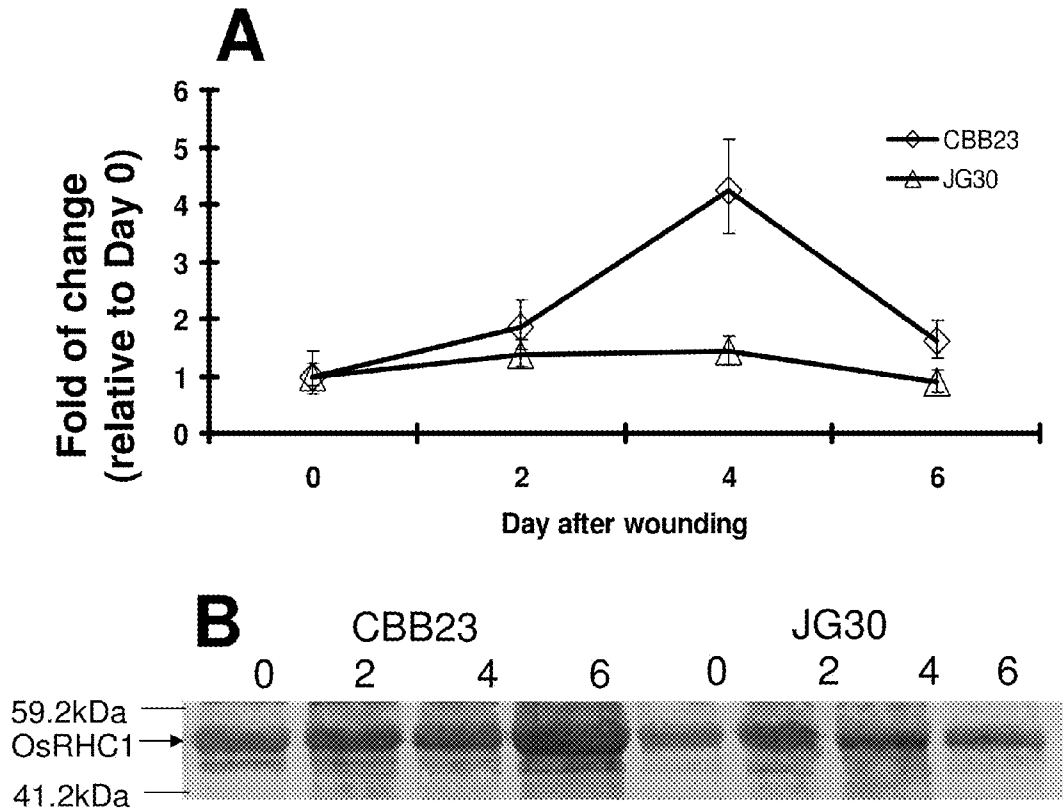


Figure 4

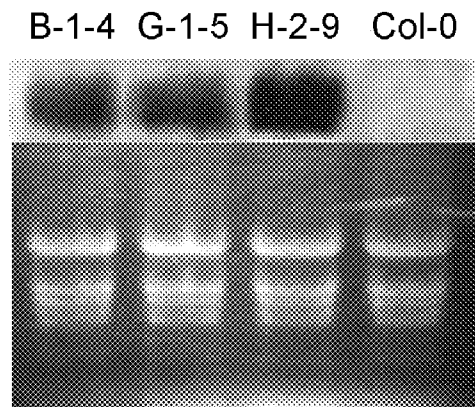


Figure 5A

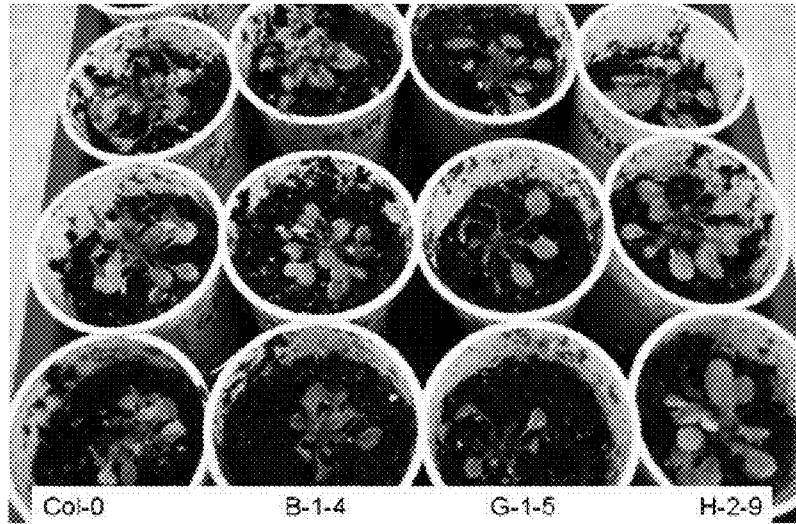


Figure 5B

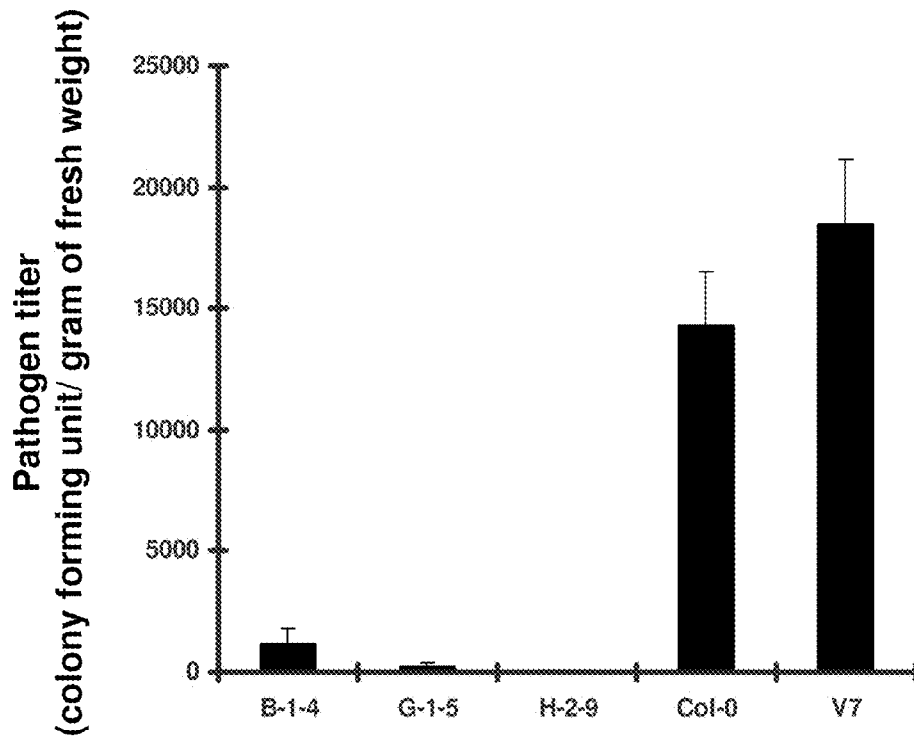


Figure 5C

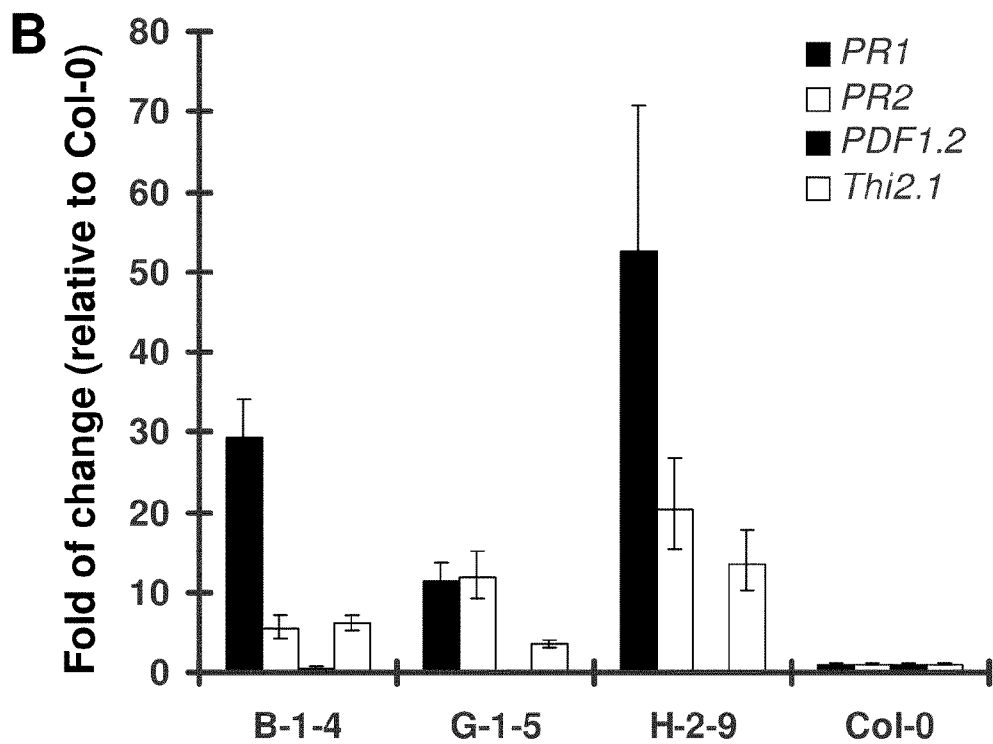
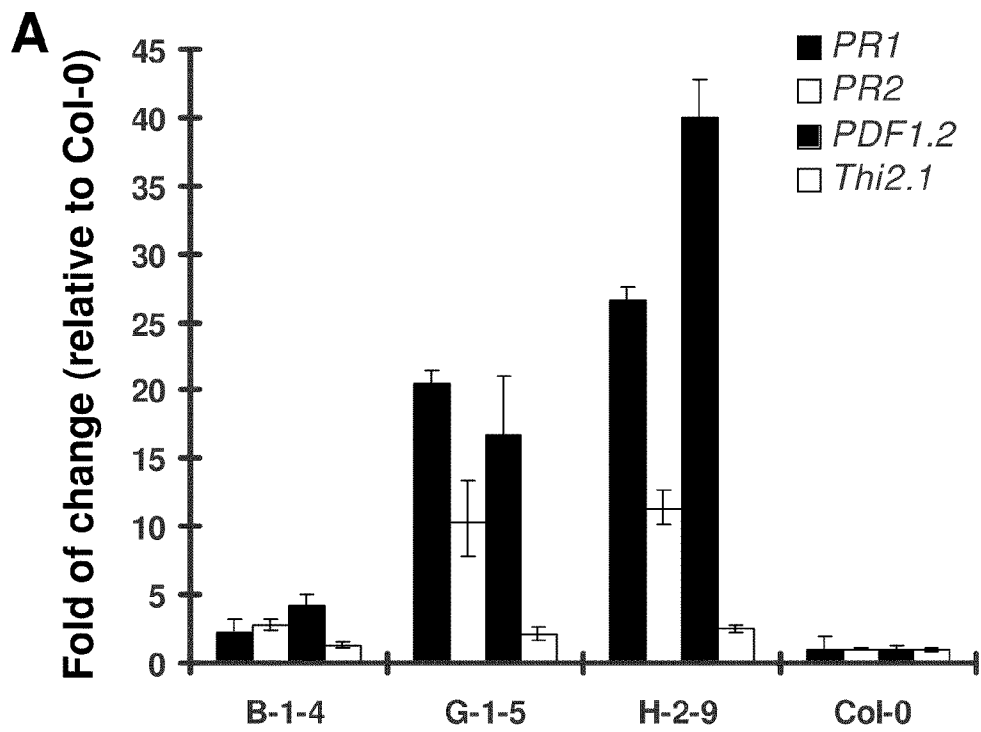
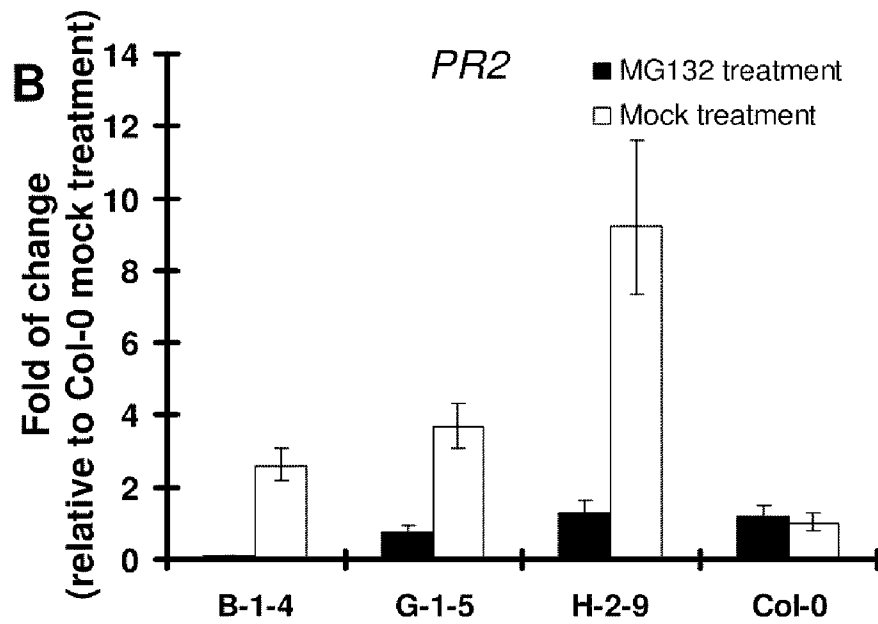
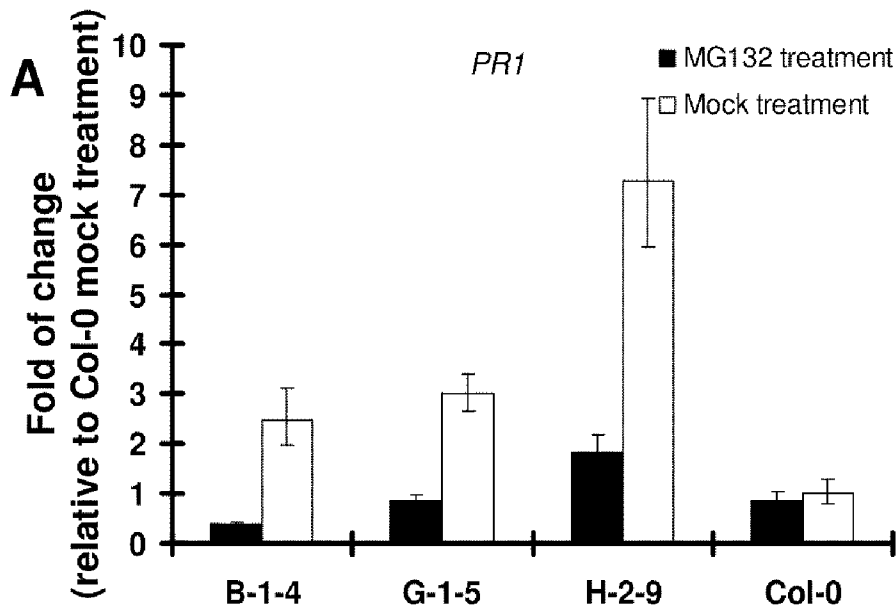
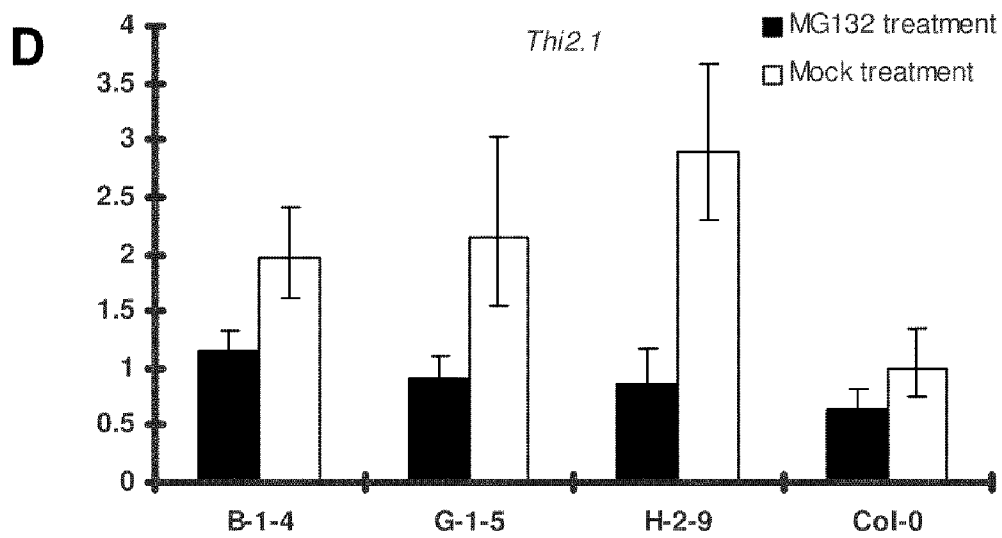
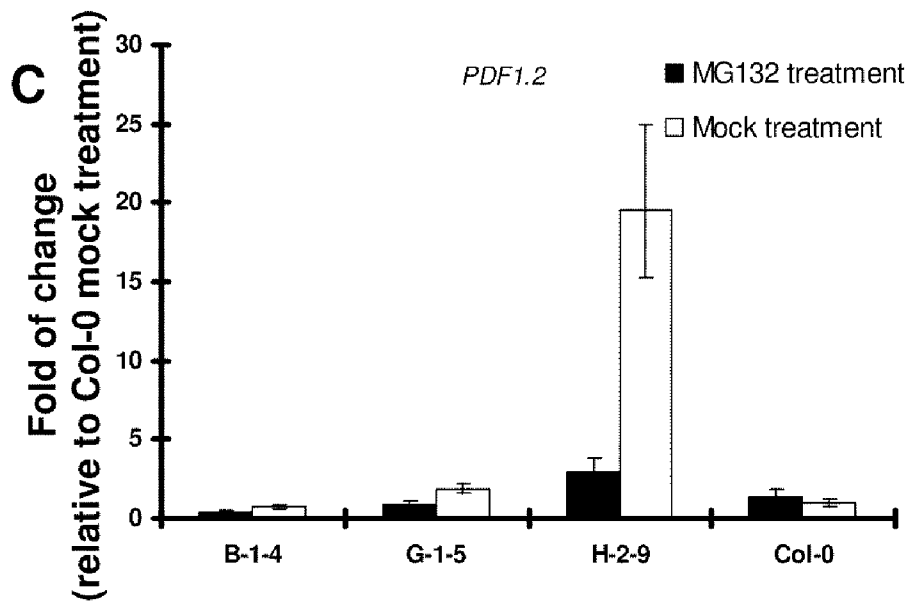


Figure 6



Figures 7A and B



Figures 7C and D

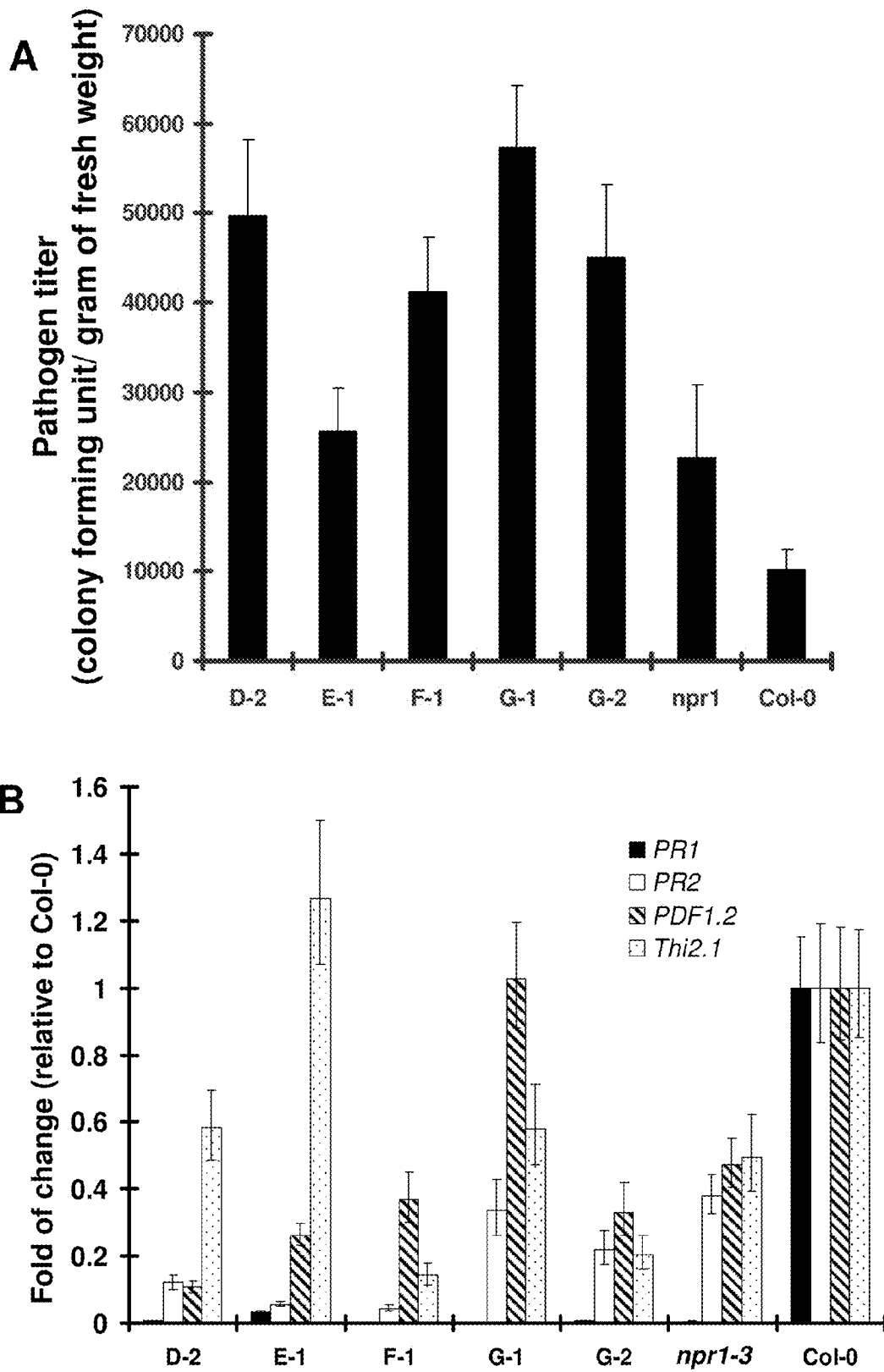


Figure 8

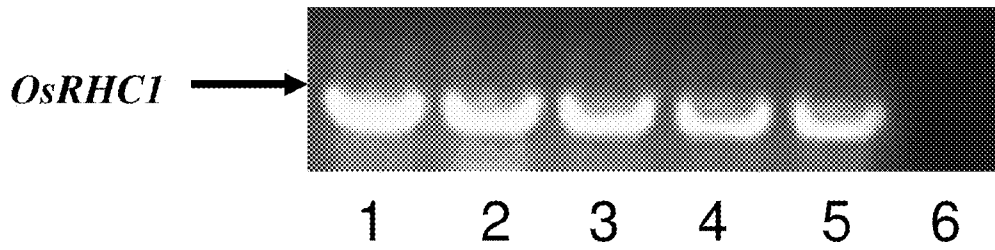


Figure 9

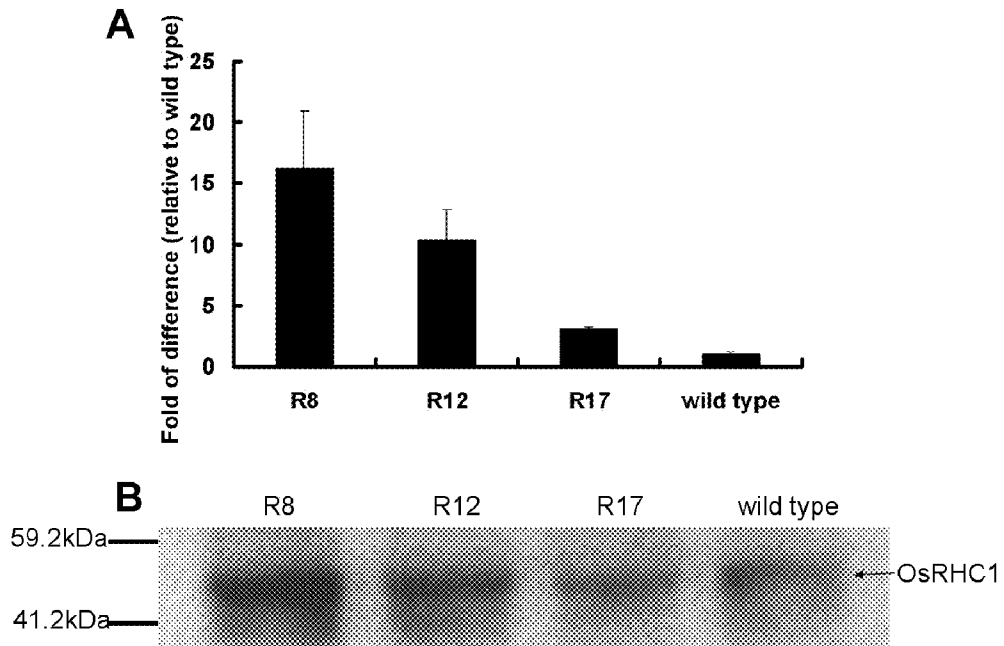


Figure 10

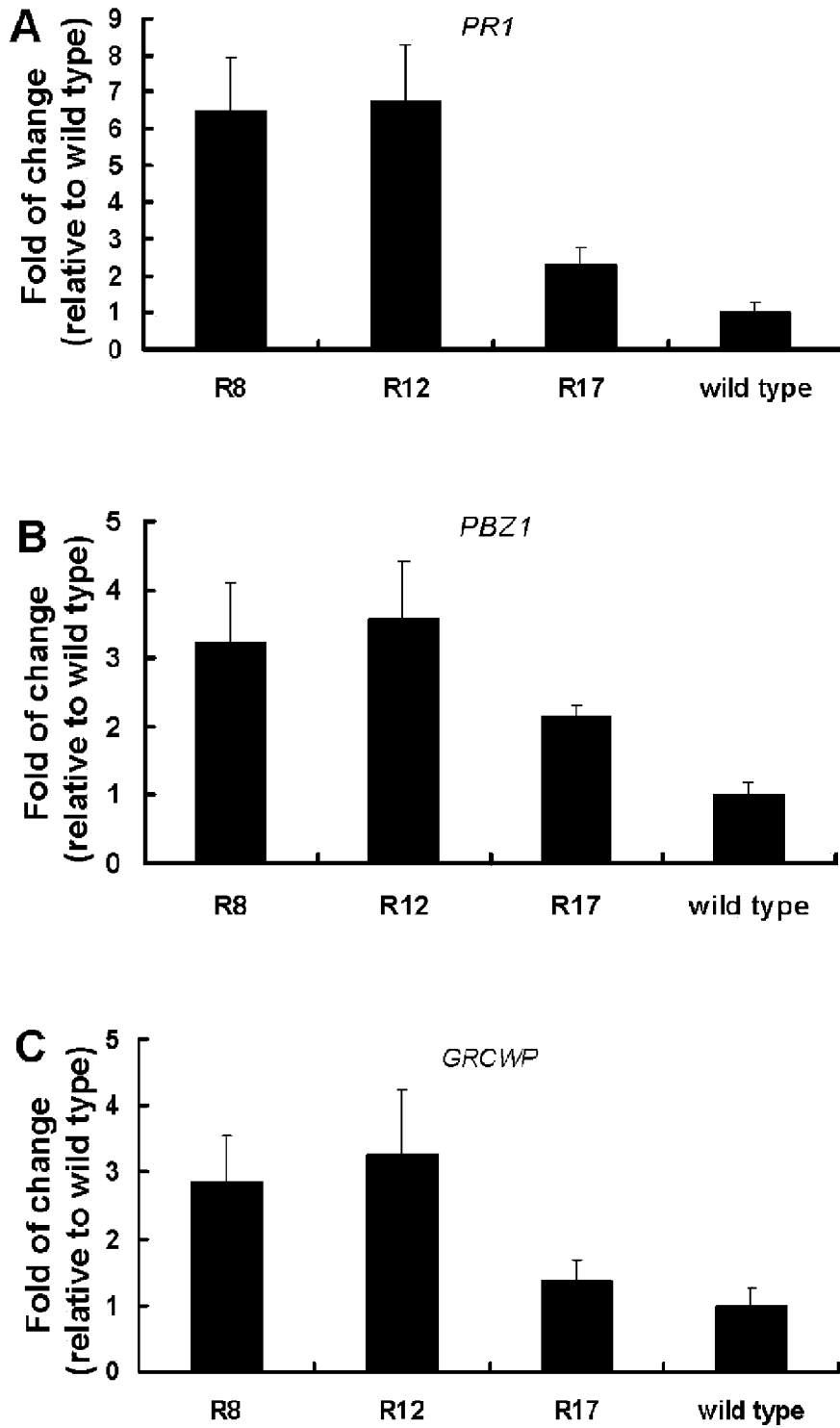


Figure 11

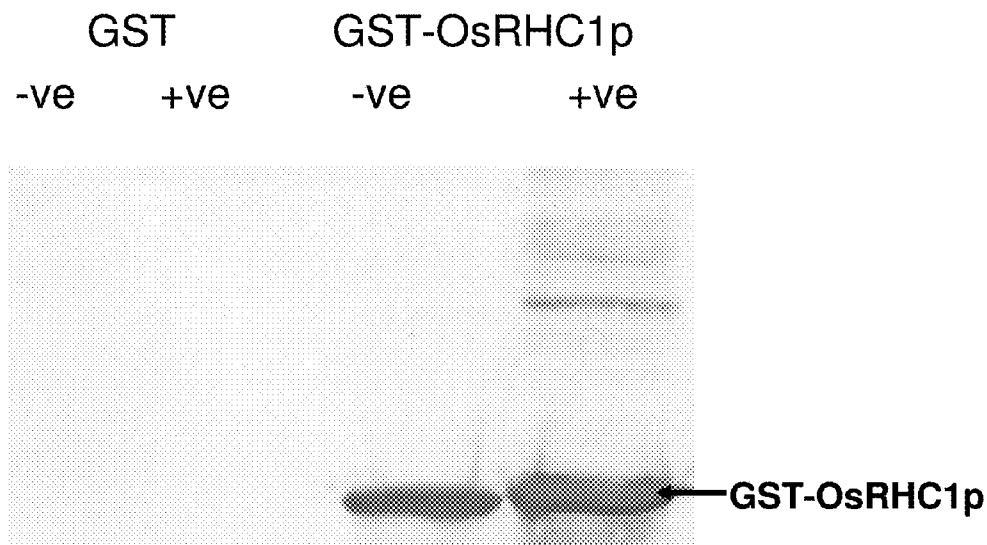


Figure 12

```

atggccgtgggggtcagagcggctcggcgaggaggcccccggcggcagctcggcgaggca
M A V G S E R L G E E A A R R Q L G E A
aggaaggccagaggcggctgctcggcgacgagggacggcgccgatgatgagggccggcgg
R K A R G G C S A T R D G A D D E G R R
cagataaacctccctcccccggigtggtcgteccctccctcactccctcttctctcaga
Q I N P P S P V W S S P P S L P L P L R
tctgcccggagggggacgggtggagggccggcgccctccctccctcttctctcagatc
S A R R G T G G G R R P P F P L S S Q I
cgcccgggtggggagagggccaccggcggcagcggcatggccctccctctgcagcagtaga
R P V G R G H R R Q R H G P P L C S S R
ggcgggcagggaggaggccacagagctgtgtttttttatttggtttttatttttga
G R Q G G G H R A V F F Y L F L F -
    
```

Figure 13

METHOD TO IMPROVE PLANT RESISTANCE TO INFECTIONS

RELATED APPLICATIONS

This application claims priority from U.S. provisional applications 60/947,590 filed 2 Jul. 2007 and 60/947,365 filed 29 Jun. 2007. The contents of these applications are incorporated herein by reference in their entirety.

REFERENCE TO SEQUENCE LISTING SUBMITTED VIA EFS-WEB

The entire content of the following electronic submission of the sequence listing via the USPTO EFS-WEB server, as authorized and set forth in MPEP §1730 II.B.2(a)(C), is incorporated herein by reference in its entirety for all purposes. The sequence listing is identified on the electronically filed text file as follows:

File Name	Date of Creation	Size (bytes)
549072000700Seqlist.txt	Jul. 23, 2010	51,884 bytes

TECHNICAL FIELD

The invention relates to proteins that improve the resistance of plants to infections, including infections by pathogen and wounding. The invention also concerns methods to improve the resistance of plants to infections by effecting expression of the genes encoding these proteins.

BACKGROUND ART

Preformed and induced defense mechanisms provide a wide spectrum of resistance toward numerous pathogens encountered by the plant host. Pathogen specific defense responses are usually initiated by the recognition of a pathogen avirulent (Avr) protein by the corresponding resistance (R) protein of the host. Ultimately, the plant host will produce a series of defense molecules (including pathogenesis-related proteins) to restrict or kill the pathogens. The processes between the initiation of resistance and the production of resistance proteins involve a complex signal transduction network which is yet to be fully elucidated.

In *Arabidopsis thaliana*, many important hubs of the defense signaling network have been identified by molecular genetic approaches, including EDS1 (Enhanced Disease Susceptibility 1), NPR1 (Non-Expresser of PR Genes 1) and NDR1 (Non Race-Specific Disease Resistance 1). Using similar tactics and together with biochemical studies, the involvement of phytohormone signals in defense responses has been corroborated in *A. thaliana*, especially the roles of salicylic acid (SA), and the other phytohormones such as jasmonic acid (JA) and ethylene (ET).

Many known signaling strategies are employed in plant defense responses. For instance, some R proteins are receptor kinases while other protein kinases also play significant roles. Biochemical signals such as calcium flux and oxidative burst are also important. Furthermore, there are several reports on the participation of other signaling components such as G-proteins and RING (Really Interesting New Gene) zinc finger proteins.

RING zinc finger proteins are a group of diverse proteins with highly conserved zinc binding domains. Based on the type of cysteine (C) and histidine (H) residue combination, the RING zinc finger domain can be classified into canonical and modified RING zinc fingers. The canonical RING zinc finger can be further grouped into two subclasses: HC subclass (consensus: C—X₂—C—X₉₋₃₉—C—X₁₋₃—H—X₂₋₃—C—X₂—C—X₄₋₄₈—C—X₂—C) (SEQ ID NO:1) and H2 subclass (consensus: C—X₂—C—X₉₋₃₉—C—X₁₋₃—H—X₂₋₃—H—X₂—C—X₄₋₄₈—C—X₂—C) (SEQ ID NO:2) (Stone, S. L., et al., *Plant Physiology* (2005) 137:13-30). Modified RING zinc fingers include RING-C2, RING-v, RING-D, RING-S/T and RING-G.

Many members of the RING zinc finger protein family (including both HC and H2 subclasses) are E3 ubiquitin ligases. Different subclasses of the RING zinc finger domain determine specificity toward different E2 ubiquitin conjugating enzymes. Other RING zinc finger proteins can bind to nucleic acids or interact with other protein targets. Besides the ubiquitin mediated degradation pathway, RING zinc finger proteins also play important roles in organelle transport and transcription/translation regulations.

In rice, more than 30 resistance loci (Xa loci) against the pathogen *Xanthomonas oryzae* pv. *oryzae* (Xoo) have been identified and 6 Xa genes were cloned mainly by map-based cloning approaches. Several pathogenesis-related (PR) genes have been reported to contribute directly to the resistance mechanism. However, only a few key components of the signal transduction pathway from the onset of R protein-Avr protein interaction to the actual resistance development have been studied. To obtain new signal transduction components related to Xoo resistance in rice, cDNA clones differentially expressed in rice lines harboring Xa loci were searched for.

The present inventors have cloned and characterized a novel RING zinc finger protein gene (OsRHC1) from rice. OsRHC1 is differentially expressed under wounding in near isogenic lines containing the Xa14 or Xa23 resistance loci, but not in the corresponding susceptible recurrent parents. Ectopic expression of OsRHC1 in transgenic *A. thaliana* enhances its resistance toward bacterial pathogens and such protective function depends on the action of the 26S proteasome.

DISCLOSURE OF THE INVENTION

A variety of genes encoding infection resistance proteins is known in plants, and various transgenic plants modified to produce them have been used in attempts to confer resistance to infections. However, these resistance proteins appear to have a limited spectrum of activity with respect to the types of pathogens that they will successfully recognize. Many cause negative side effects (such as programmed cell death) as well. The present invention provides materials that can be used to confer resistance to infections on a wide variety of plants, without apparent negative side effects. The invention provides recombinant materials for the production of a protein designated OsRHC1 which is a RING zinc finger protein that confers resistance to infections of a broad spectrum of pathogens. Because the protein of the invention which is derived from a monocot (rice) is also effective in dicots (*Arabidopsis*) it is applicable to a broad spectrum of plants as well.

In one aspect, the invention is directed to expression systems that produce the OsRHC1 protein and proteins closely related thereto that are RING zinc finger proteins and are able to improve resistance of plants to infections. Transgenic plants modified with the expression systems of the invention

have enhanced ability to resist infections either from pathogenic organisms or by wounding.

Thus, in another aspect, the invention is directed to plant cells or plants that have been modified to contain an expression system that produces this RING zinc finger protein. The plants may either be heterologous from the origin of OsRHC1 or may be rice plants modified to overexpress this protein.

In still another aspect, the protein produced by this expression system may be used to conduct screening assays to identify compounds or combinations of compounds that modulate resistance to infections in plants.

The invention also relates to antibodies that are immunospecific for the OsRHC1 protein. These antibodies are useful for detecting and purifying this protein.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows the nucleotide sequence-encoding region of the OsRHC1 gene and the amino acid sequence of the OsRHC1 protein (SEQ ID NOS:42-43).

FIG. 2A shows the full-length amino acid sequence of OsRHC1 (SEQ ID NO:43) aligned to seven annotated proteins (SEQ ID NOS:44-50) exhibiting high degree of similarity. FIG. 2B shows membrane bound and soluble protein fractions extracted from CBB23 and JG30 followed by Western blot analysis using anti-OsRHC1 antibodies.

FIGS. 3A and 3B are graphs showing expression of OsRHC1 in bacterial blight resistant lines CBB14 and CBB23 (carrying the Xa14 locus and Xa23 locus, respectively) and their susceptible recurrent parents (SN1033 and JG30, respectively).

FIG. 4A is a graph showing wounding-induced expression of OsRHC1 by real-time PCR. FIG. 4B shows a Western blot of the corresponding protein.

FIGS. 5A-C show pathogen inoculation tests of transgenic *A. thaliana* expressing OsRHC1. The expression of the transgene OsRHC1 in the transgenic lines was confirmed by Northern blot analysis in FIG. 5A. The disease symptoms were visible as shown in FIG. 5B and the rosette leaves (not at the site of infection) were harvested to estimate the titer of pathogens shown in FIG. 5C.

FIGS. 6A and 6B show expression of defense marker genes in transgenic *Arabidopsis thaliana* without (A) and with (B) *Pseudomonas syringae* pv. *tomato* DC3000 (Pst DC3000) inoculation.

FIGS. 7A-D are graphs showing expression of defense marker genes (PR1 (A), PR2 (B), PDF1.2 (C) and Thi2.1 (D)) when treated with MG132 (a 26S proteasome inhibitor).

FIGS. 8A and 8B show the results of pathogen inoculation test of OsRHC1 transgenic *A. thaliana* in the npr1-3 background. FIG. 8A shows the expression of the OsRHC1 gene and FIG. 8B shows the expression of four defense marker genes.

FIG. 9 shows the results of PCR screening of the OsRHC1 transgene in transgenic rice lines.

FIG. 10A shows the expression of OsRHC1 by real-time PCR and FIG. 10B shows production of the corresponding protein in transgenic rice lines.

FIGS. 11A-C show expression of defense marker genes PR1 (A), PBZ1 (B) and GRCWP (C) in OsRHC1 transgenic rice lines.

FIG. 12 shows the results of autoubiquitination assay conducted on the RING-HC-C-terminal portion of OsRHC1.

FIG. 13 shows the DNA sequence (SEQ ID NO:51) and deduced amino acid sequence (SEQ ID NO:52) of a binding partner for OsRHC1.

MODES OF CARRYING OUT THE INVENTION

A protein designated rice RING-HC subclass protein-1 (OsRHC1) is a 409-amino acid protein overexpressed in rice in response to pathogen or wound-induced infections. This protein and its variants, which share at least 90%, preferably 95%, more preferably 98% or 99% sequence identity over the entire length of this 409-amino acid sequence (shown in FIG. 1) are able to confer resistance to the negative effects of infection to a wide variety of plants when said plants are modified to produce these proteins (collectively referred to as OsRHC1 proteins). The present invention provides expression systems that can be used to modify a wide variety of plants, both monocots and dicots, to enhance their ability to resist infections. The generic capability of such expression systems to confer resistance is confirmed in the examples hereinbelow which demonstrate that the protein, which has its origin in the monocot, rice, is able to confer these properties on the dicot *A. thaliana*.

The techniques for constructing expression vectors operable in plants, for modifying plant cells, for regenerating plant cells into intact plants and recombinant manipulation of plants in general are by this time well known. A summary of such techniques is found, for example, in U.S. Pat. No. 7,109,033 which is incorporated herein by reference for its disclosure of these techniques.

As noted in this patent, promoters useful in plant expression may be constitutive, inducible and/or tissue-specific. Transformation techniques include use of *Agrobacterium*, lipofection, electroporation, and the like. Techniques for regeneration of plants from transformed plant cells are also well established.

Accordingly, once the nucleotide sequence encoding the OsRHC1 protein is available, methods of preparing transgenic plants that produce these proteins are well within the ordinary skill of the art. The nucleotide sequence natively producing this protein has been deposited in GenBank with Accession No. EF584506 and synthetic alternatives having variations in codon usage are possible.

Thus, according to the invention, a suitable expression system is constructed for operability in plants wherein the nucleotide sequence encoding the proteins of the invention is operably linked to suitable control sequences operable in plants. This expression system is used to modify plant cells or plants so that the protein is produced either ubiquitously in plant tissues or in specialized desired locations in the plant, depending on the choice of control system and method of transformation. The resulting plants, whether monocots or dicots, are then permitted to produce the protein in response to pathogen or wound-induced infection so as to enhance their ability to resist damage caused by these infectious events.

As shown below, OsRHC1 is an E3 ubiquitin ligase which enhances the destruction of unwanted proteins by directing them to the proteasome. This property is shared in common with other RING proteins, and represents one aspect of its protective function. This protein is the first E3 that harbors transmembrane domains at the N-terminal region and RING-HC at the C-terminal cytoplasmic tail that has been found to be involved in plant disease resistance.

In addition, the protein itself, produced in sufficient quantity and isolated and purified to a suitable extent (at least 50% pure by weight, preferably 75% pure, more preferably 90% or 95% pure) can be used as a screening tool. Compounds or combinations of compounds that are able to bind the protein are candidates for modulating the ability of plants to resist infection. Compounds or combinations of compounds that,

by binding the protein, are able to agonize its activity will enhance the infection-resisting capability of plants that are able to produce this protein.

Thus, the binding assay may be used as a preliminary screen. As it has been shown that OsRHC1 is an E3 ubiquitin ligase, the assay for ubiquitin ligase activity described below in Example 9, or a similar assay for such activity is used to demonstrate the agonist activity of a candidate compound. Thus, a suitable candidate will enhance the ability of OsRHC1 to effect ubiquitination.

Also useful for purifying the proteins of the invention and for detecting them are antibodies that are specifically immunoreactive with said proteins. The term "antibodies" is understood to mean complete antibodies, polyclonal or monoclonal, as well as the immunospecific fragments thereof such as Fab fragments, as well as recombinantly produced forms such as single-chain F₂ antibodies. Thus, the term "antibodies" refers both to any form of the antibody and to any portion thereof that retains its immunospecific characteristics. Such antibodies can be used, for example, on affinity columns, etc., for purification.

In the examples below, the nucleotide sequence encoding the OsRHC1 protein shown in FIG. 1 has been retrieved from rice and deposited. Further, it was demonstrated that the OsRHC1 expression, both at an mRNA level and at a protein level, could be induced in a line of rice that exhibits resistance in response to a pathogen and in response to wounding.

Transgenic *A. thaliana* plants were obtained using an expression construct for the OsRHC1 protein, and these transgenic plants were shown to have enhanced expression of four defense marker genes, both under regular growth conditions and when salicylic acid or jasmonic acid was added. The OsRHC1 transgenic *A. thaliana* also showed constitutive expression of the OsRHC1-encoding DNA and was protected by this expression when challenged with a *Pseudomonas*. Similarly, overexpression of this DNA in rice resulted in expression of several defense marker genes.

The following examples are offered to illustrate but not to limit the invention.

EXAMPLE 1

Identification and Cloning of OsRHC1-Encoding cDNA

One partial cDNA clone was obtained via suppression subtractive hybridization techniques with the PCR-select cDNA subtraction kit (Clontech 637401), using total RNA extracted from six to eight-week-old CBB14 which is bacterial blight resistant (tester) and SN1033 the susceptible parent of CBB14 (driver) rice lines collected four days after pathogen (Xoo race LN44) inoculation. Inoculation was performed by clipping method described in Zhang, Q., et al., *Acta Agr. Sin.* (1996) 22:135-141. Using the DNA sequence information of this partial clone, 5'-Rapid-Amplification of cDNA Ends (5'-RACE) experiment and subsequent PCR amplifications using specific primers were performed. Gene specific primers 5'-TTCTCC ATGTTCGGTAAACCTTTC-3' (SEQ ID NO:3), 5'-TAAAGTTGTGATTGAGACTACA TGG-3' (SEQ ID NO:4) and 5'-ACATTGCACAACCAACATGTAC-3' (SEQ ID NO:5) were employed in the 5'RACE reactions. To amplify the full length coding region, PCR using the primer pair 5'-CCTCACTTTTGTCTCCAC-3' (SEQ ID NO:6) and 5'-CGACATTGCACA ACCAAC-3' (SEQ ID NO:7) were performed. All clones were stored in the plasmid vector pBluescript® KSII(+) (Stratagene) and propagated in the *E. coli* strain DH5α.

The resulting cDNA clone (GenBank accession number EF584506) encodes an intact open reading frame of 409 amino acid residues (FIG. 1). EF584506 is 99% identical to a directly deposited rice cDNA clone (accession number: NM_001057564). The corresponding gene in the rice genome appears to be a single copy gene located on chromosome 3. BlastP search showed that the protein encoded by our clone exhibits 99% identity to a rice clone annotated as a zinc finger family protein (accession number: ABF98464), but missing 64 amino acid residues at the N-terminus. Further analysis using the conserved domain database (CDD) revealed that the predicted protein harbors a RING zinc finger domain. The pattern of the conserved cysteine and histidine residues in the RING zinc finger domain exhibited a signature for the RING-HC subclass. The clone was designated as OsRHC1 accordingly.

The predicted amino acid sequence of the OsRHC1 protein was compared with two RING zinc finger proteins, EL5 (RING-H2 subclass) and XB3 (RING-HC subclass) from rice that are involved in disease resistance. No significant homology was found except at the RING zinc finger domain (data not shown). The RING zinc finger domain of OsRHC1 is located at the C-terminus (FIG. 2A) while such domain in EL5 and XB3 is located in the middle portion or close to the C-terminus of the protein, respectively. Prediction by the TopPred and the iPSORT programs suggested that OsRHC1 may possess multiple transmembrane domains (FIG. 2A) while EL5 only has one and XB3 does not possess any transmembrane region with high certainty (data not shown).

BlastP analysis revealed that OsRHC1 shares high amino acid sequence homology to seven other annotated proteins deposited in GenBank from various plant species (FIG. 2A). These proteins exhibit greater than 50% identity (spanning full length) to OsRHC1, with multiple transmembrane domains at the N terminal half, and a RING-HC domain at the C-terminus. The consensus of the RING-HC domain for this group of proteins is Cys-X₂-Cys-X₁₁-Cys-X-His-X₃-Cys-X₂-Cys-X₆-Cys-X₂-Cys (SEQ ID NO:8). There is apparently no published information on the functions of these homologues.

EXAMPLE 2

Demonstration that OsRHC1 is Membrane Bound

To verify that the OsRHC1 is membrane bound as depicted by bioinformatics tools, membrane-bound and soluble proteins were separated using a fractionation protocol (modified from Jiang and Rogers, *J. Cell Biol.* (1998) 143:1183-1199). For Western blot analysis, the proteins were electrophoretically separated on a polyacrylamide gel (4% stacking; 10% resolving) before transferred to an activated PVDF membrane (pre-treated in absolute methanol for 20 minutes followed by protein transfer buffer for 15 minutes) using the Trans-Blot® SD Semi-Dry Electrophoretic Transfer Cell (Bio-Rad 170-3949). The blocking and detection steps were performed according to the manufacturer's manual (Western Breeze™ Immunodetection Kit, InvitroGen WB7106). Primary antibodies (polyclonal) targeting the OsRHC1 protein was raised by a commercial service (InvitroGen, Custom antibody) via injecting a synthetic peptide ('N'-CGYPPEV-VRKMPKRD-'C') (SEQ ID NO:9) into rabbits and antibodies were purified using affinity column before use. Anti-rabbit secondary antibody conjugated to an alkaline phosphatase (provided in Western Breeze™ Immunodetection Kit, InvitroGen WB7106) was used to recognize the primary antibody.

ies. Western blot analysis confirmed that the OsRHC1 protein was tightly associated to membranes (FIG. 2B).

EXAMPLE 3

OsRHC1 is Wound-Inducible in the Rice Lines CBB14 and CBB23

To study the expression pattern of OsRHC1, real-time PCR analyses was performed using reverse-transcribed RNA samples from two near isogenic pairs (CBB14 containing Xa14 and its susceptible recurrent parent SN1033; CBB23 a resistant line containing Xa23 and its susceptible recurrent parent JG30). Rice lines were grown on regular field soil in a green house (temperature 24-28° C.; RH 70-80%; under natural light). Inoculation of the Xoo races LN44 and P6 was performed by clipping method described in Zhang, Q., et al., supra (1996). Mock inoculation and wounding treatment followed the same procedure except that the pathogen was replaced by water. For the time-course experiments, samples were collected at 0, 2, 4, 6 days at around the same time of the day (between 8-10 am). Day 0 sample was collected before treatment.

For evaluating expression of OsRHC1 via real-time PCR, total RNA was extracted by the phenol extraction method of Ausubel, et al., *Current Protocols in Molecular Biology* (1995) J. Wiley & Sons, New York. The cDNA samples were generated by reverse transcription (18-mer oligo-dT; SUPERSCRIPTM II RNaseH (InvitroGen 18064-071)) of DNase I (InvitroGen 18068-015)-treated RNA samples.

Real-time PCR amplification of cDNA was conducted using the ABI PRISM 7700 Sequence Detection System (Applied Biosystems) in 96-wells PCR plate with dome cap. Reaction was carried out in a 20 µl reaction volume containing 10 µl SYBR Green PCR Master Mix (Applied Biosystems 4309155) with 0.3 µM each of the forward and reverse primers. OsRHC1 primers for real-time PCR were 5'-AAAGAA-GAGCAAGCCCGTTAT-3' (SEQ ID NO:10) and 5'-GC-CTCCAATACCTTCTGCAA-3' (SEQ ID NO:11). All reactions were set independently for at least four times and at least three sets of consistent data were used for analysis. The expression level of actin (*O. sativa* OsAc1D; accession number: X15865) with the primer set 5'-CTTCATAGGAATG-GAAGCTGCGGGTA-3' (SEQ ID NO:12) and 5'-GACCAC-CTTGATCTTCATGCTGCTA-3' (SEQ ID NO:13) was used to normalize the results. The relative gene expression was calculated using the 2^{-ΔΔCT} method of Livak and Schmittgen, *Methods* (2001) 25:402-408.

To validate the reliability data, amplification efficiencies between the target genes and the housekeeping genes of all the real-time PCR reactions were compared, and dissociation curves of all PCR products were examined to ensure the quality of PCR. At least two independent batches of plant samples were used and gene expression patterns were consistently observed. All PCR products were sequenced at least one time to verify that the right targets were being quantified.

When an incompatible Xoo strain (LN44 for Xa14 and P6 for Xa23) was inoculated, the rice lines containing Xa14 or Xa23 exhibited an induction of OsRHC1 gene expression while the susceptible recurrent parents were non-responsive as shown in FIGS. 3A and 3B, respectively. However, such induction was also observed in mock inoculated samples which had been wounded, suggesting that OsRHC1 could be wounding-inducible. The amplitude of induction was much stronger in the case of CBB23 which harbors the Xa23 locus that confers broad spectrum resistance.

The effect of wounding on OsRHC1 expression in CBB23 line and its susceptible recurrent parent JG30 was further analyzed. Both RNA and protein samples were collected after wounding by leaf clipping. CBB23 and JG30 rice lines (eight-week-old plants) were wounded by clipping. Day 0 leaf samples were collected before wounding. Leaf tissues about 6-8 mm away from the wounding site were collected at 2, 4, and 6 days after clipping. Total RNA and membrane bound protein samples were prepared in parallel. Real-time PCR experiments were performed as described above. Western blot analysis was performed using the anti-OsRHC1 antibody as described in Example 2.

The induction peak of OsRHC1 gene expression appeared on Day 4 after treatment in CBB23 (FIG. 4A). Western blot analysis of membrane-bound proteins showed that the production of the OsRHC1 protein in CBB23 was greatly enhanced on Day 6 (FIG. 4B), after the induction of gene expression on Day 4. The response in JG30 was not obvious when compared to CBB23, indicating that the presence of the Xa23 locus may play a role in the wounding induction of OsRHC1.

EXAMPLE 4

Production of Transgenic *Arabidopsis* Lines

To test whether OsRHC1 could mediate resistance in dicots, an *A. thaliana* was modified to produce OsRHC1 protein and challenged with *Pseudomonas syringae* pv *tomato* DC3000. OsRHC1 cDNA was inserted into a binary vector and the transgene expression was driven by the Cauliflower Mosaic Virus 35S promoter. *Agrobacterium*-mediated transformation of the wildtype Col-0 *A. thaliana* line was performed using a vacuum infiltration method (Bechtold, N., et al., *Methods Mol. Biol.* (1998) 82:259-266). Transgenic plants with single insertion locus were screened by kanamycin resistance phenotype (encoded by the selection marker gene in the binary vector) of offspring. A 3:1 (resistant:sensitive) ratio verified by Chi-Square test in the T1 generation suggested a single insertion event.

Only positive transformants containing a single insertion locus were propagated to obtain homozygous lines for further experiments. The transgene expression in three independent homozygous transgenic lines was examined by Northern blot analysis. As shown in FIG. 5A, three transformed lines, B-1-4, G-1-5, and H-2-9, showed high levels of production of mRNA. However, the Col-0 line, the wildtype, showed no production of the mRNA.

A. thaliana was grown in a growth chamber (temperature 22-24° C.; RH 70-80%; light intensity 80-120 µE of a 16 h light-8 h dark cycle). The preparation of the Pst DC3000 culture, inoculation (by a dipping method), and subsequent titering were performed as previously described (modified from Kim, H. S., et al., *Plant Cell* (2002) 14:1469-1482; Uknes, S., et al., *Plant Cell* (1992) 4:645-656). Six-week-old seedlings were challenged with Pst DC3000 in a concentration of 10⁸ colony forming unit/ml in 10 mM MgSO₄ supplemented with 0.02% (v/v) Silwet L-77 (Pieterse, C. M. J., et al., *Plant Cell* (1998) 10:1571-1580; Ton, J., et al., *Mol. Plant-Microbe Interact.* (2002) 15:27-34).

Pst DC3000 inoculation caused severe yellowing and necrosis in infected Col-0 and transgenic plants transformed with the empty vector V7, while the disease symptoms were much reduced in all OsRHC1 transgenic lines as shown in FIG. 5B.

The titers of pathogen inside the rosette leaves were consistent with the observed phenotypes (FIG. 5C). Furthermore,

the H-2-9 line that exhibited the highest level of transgene expression also gave the lowest pathogen titer (comparing FIGS. 5A and 5C).

EXAMPLE 5

Expression of OsRHC1 Enhances the Expression of Defense Marker Genes in Transgenic *Arabidopsis thaliana*

The expression of four defense marker genes, PR1, PR2, PDF1.2 and Thi2.1, was tested in transgenic *A. thaliana*. In *A. thaliana*, these genes are indicators of defense pathways mediated by different phytohormones including SA, JA, and ET.

Leaf tissues of six-week-old *A. thaliana* transgenic lines (B-1-4, G-1-5 and H-2-9) expressing OsRHC1 and the untransformed wild-type (Col-0) were harvested to prepare total RNA, followed by real-time PCR as described in Example 3. The primers used are as follows:

PR1:
5' - TCAAGATAGCCACAAGATTATC - 3' (SEQ ID NO: 14)
and
5' - CTTCTCGTTCACATAATTCCAC - 3' ; (SEQ ID NO: 15)

PR2:
5' - ACCACCACTGATACGTCCTC - 3' (SEQ ID NO: 16)
and
5' - AACTTCATACTTAGACTGTCGATC - 3' ; (SEQ ID NO: 17)

PDF1.2:
5' - CCCTTATCTTCGCTGCTCTTGT - 3' (SEQ ID NO: 18)
and
5' - CCCTGACCATGTCCCCTTG - 3' ; (SEQ ID NO: 19)

Thi2.1:
5' - AGCACTGCAAGTTAGGGTGTGA - 3' (SEQ ID NO: 20)
and
5' - ACATTGTTCCGACGCTCCAT - 3' . (SEQ ID NO: 21)

The tubulin (*A. thaliana* β -tubulin 4, accession number: M21415) with the primer set 5'-GAAGGTGCTGAGT-TGATTG-3' (SEQ ID NO:22) and 5'-GGACTTGACGT-TGTTTGG-3' (SEQ ID NO:23) was used to normalize the results.

The expressions of PR1 (solid), PR2 (open), PDF1.2 (hatched), and Thi2.1 (dotted) in each transgenic line as shown in FIG. 6A were compared to those of Col-0 (expression level set to 1).

In six-week-old seedlings under regular growth conditions, all four defense marker genes exhibited enhanced expression when compared to the wild type Col-0 (FIG. 6A). The fold of induction was particularly higher for the PR1 and the PDF1.2 genes which are mediated by two different signaling pathways. Among three independent transgenic lines tested, the H-2-9 line that showed highest expression of the transgene and best protection in the pathogen inoculation experiment also gave the highest fold of induction of PR1 and PDF1.2 (comparing FIGS. 5 and 6A).

When the plants were subjected to the challenge of Pst DC3000, the levels of PR1 and PR2 transcripts in Col-0 increased (data not shown) but the expression levels of these genes were even higher in transgenic lines (FIG. 6B). While the level of Thi2.1 in Col-0 did not alter significantly by the pathogen inoculation (data not shown), its expression was elevated in the transgenic lines (FIG. 6B). The expression of PDF1.2, on the other hand, was repressed by Pst DC3000 inoculation in both Col-0 and transgenic lines (FIG. 6B).

To show the relationship between the function of OsRHC1 and ubiquitin-mediated protein degradation, the effects of MG132 (a 26S proteasome inhibitor) on the expression of defense marker genes were studied in the transgenic lines. Four-week-old seedlings were subjected to MG132 treatment. The 26S proteasome inhibitor (MG132) was applied to the transgenic *A. thaliana* using a protocol modified from previous reports (Abas, L., et al., *Nature Cell Biol.* (2006) 8:249-256; Dong, C. H., et al., *Proc. Nat'l Acad. Sci. USA* (2006) 103:8281-8286; Guo, H., et al., *Cell* (2003) 115:667-677; Oñate-Sánchez, L., et al., *Plant Physiol.* (2002) 128: 1313-1322). In brief, 50 mg/L MG132 dissolved in 1% (v/v) DMSO supplemented with 0.01% (v/v) Silwet L-77 were poured onto MS square plates to cover the roots but not the aerial tissues of the seedlings. Mock treatment was performed with the same procedures except that no MG132 was added. After four hours, the seedlings were harvested for RNA extraction followed by real-time PCR.

Expression of the transgene was not affected by such treatment (data not shown). In Col-0, no significant effects of MG132 on the expression of defense marker genes were observed. On the other hand, the induction effects of overexpressing OsRHC1 on the four defense marker genes were diminished under MG132 treatment, as shown in FIGS. 7A-7D for PR1, PR2, PDF1.2, and Thi2.1, respectively, where open bars represent mock treatment and solid bars represent MG132 treatment

In summary, it appears that the ability of the OsRHC1 protein to enhance the expression of the defense marker genes was, in all cases, inhibited by an inhibitor of the 26S proteasome. It thus appears that the ability of the invention protein to enhance expression of the defense marker genes may be dependent on the 26S proteasomal activity.

EXAMPLE 6

The Protective Function of the OsRHC1 Clone in Transgenic *Arabidopsis thaliana* is Dependent on the Function of NPR1

The function of OsRHC1 in relation to a known hub in the defense signaling network was positioned using the model plant system. NPR1 which mediates both SA and JA/ET signals and plays a central role in defense signaling in *A. thaliana*. OsRHC1 was transformed as described above into the *npr1-3 A. thaliana* mutant that is depleted of NPR1. Independent transformants with a single insertion locus were selected. At the time of inoculation, the expression of transgene (under the control of the Cauliflower Mosaic Virus 35S promoter) in individual lines was examined with real-time PCR as described in Example 3. The steady-state level of OsRHC1 in an *npr1-3* background was found to be comparable to that in the transgenic lines with a Col-0 background (data not shown).

Eight-week-old transgenic lines (D-2, E-1, F-1, G-1 and G-2) expressing OsRHC1, the untransformed *npr1-3* mutant, and the wild-type Col-0 were challenged by Pst DC3000 and the subsequent estimation of pathogen titer was obtained as shown in FIG. 8A. Expression of defense marker genes was determined as described in Example 5. The expressions of PR1 (solid), PR2 (open), PDF1.2 (hatched), and Thi2.1 (dotted) in each line was compared to those of Col-0 (expression level set to 1), as shown in FIG. 8B. No significant increase in the expression of four selected defense marker genes was found in any of these transgenic lines.

When the *npr1-3* transgenic lines were subjected to the challenge of Pst DC3000, no protection effects could be observed in the transgenic lines. Both the disease symptom

11

development (data not shown) and pathogen titer of these transgenic lines resembled that of the untransformed npr1-3 mutant. Thus, protection appears to require NPR1.

EXAMPLE 7

Construction of OsRHC1 Transgenic Rice

The nucleotide sequence of FIG. 1 encoding OsRHC1 was subcloned into a double T-DNA binary vector, pSB130 (from Dr. Liu Qiaoquan and Prof. Samuel Sun at the Chinese University of Hong Kong). The vector pSB130 carries two T-DNA. One T-DNA harbors the hygromycin resistance gene (selectable marker) and the other possesses a multiple cloning site downstream from a maize ubiquitin promoter for cloning of target genes. The recombinant construct was transformed into the *Agrobacterium* EHA105 for rice transformation, and transgenic rice lines were constructed.

FIG. 9 shows PCR screening of the OsRHC1 transgenes in T2 transgenic rice lines (parent: Aichi Asahi). The forward and reverse primers for PCR are from the maize ubiquitin promoter and the OsRHC1 coding region respectively as follows:

Forward primer:
5'-CTGATGCATATACATGATGG-3'; (SEQ ID NO: 24)

Reverse primer:
5'-ACATTGCACAACCAACATGTAC-3'. (SEQ ID NO: 25)

A total of five OsRHC1 transgenic rice lines were obtained.

EXAMPLE 8

Over-Expression of OsRHC1 and Defense Marker Genes in Rice

The expression of the OsRHC1 and three rice defense marker genes (PR1, glycine rich cell wall protein encoding gene (GRCWP) and PBZ1) was studied via real-time PCR as described in Example 3. PR1 is a well known PR protein. Glycine rich cell wall protein (encoded by GRCWP) is a structural protein commonly found in strengthened cell wall to hinder pathogen attacks. PBZ1 is induced by probenazole (PBZ), N-cyanomethyl-2-chloro-isonicotinamide (compounds known to induce disease resistance) as well as the fungal blast pathogen *M. grisea*. PBZ1 is induced faster by incompatible strains of *M. grisea* than compatible strains. PR1 and PBZ1 are induced by over-expression of NH1, a key signaling component in rice defense response.

The RNA was extracted from 8-week-old plants of the transgenic rice lines (at the T3 generation) carrying a single insertion of OsRHC1 and their wild type parent (Aichi Asahi). The primers used in real-time PCR are as follows:

O. sativa OsRHC1 forward primer:
5'-AAAGAAGAGCAAGCCCGTTAT-3'; (SEQ ID NO: 26)

O. sativa OsRHC1 reverse primer:
5'-GCCTCCATACCTCTTCTGCAA-3'; (SEQ ID NO: 27)

O. sativa PR1 (BF889437) forward primer:
5'-CGGACAGAGGCCCTTACTAAGTTATTT-3'; (SEQ ID NO: 28)

O. sativa PR1 (BF889437) reverse primer:
5'-GACCTGTTTACATTTTCACGTCCTTTATT-3'; (SEQ ID NO: 29)

12

-continued

O. sativa GRCWP (BF889438) forward primer:
5'-GAGGCAACGGACACCCTAAG-3'; (SEQ ID NO: 30)

O. sativa GRCWP (BF889438) reverse primer:
5'-TGTAAGCAGAGAGAGAGGCTCATT-3'; (SEQ ID NO: 31)

O. sativa PBZ1 (D38170) forward primer:
5'-AAGCTCAAGTCACACTCGAC-3'; (SEQ ID NO: 32)

O. sativa PBZ1 (D38170) reverse primer:
5'-GATGTCCTTCTCCTTCTCC-3'. (SEQ ID NO: 33)

For normalization, the actin primers are:

O. sativa OsAct1D (X15865) forward primer:
5'-CTTCATAGGAATGGAAGCTGCGGGTA-3'; (SEQ ID NO: 34)

O. sativa OsAct1D (X15865) reverse primer:
5'-GACCACCTTGATCTTCATGCTGCTA-3'. (SEQ ID NO: 35)

FIG. 10A shows the over-expression of OsRHC1 in the transgenic rice lines as measured by real-time PCR. Western blot analysis conducted as described in Example 2, gives the results shown in FIG. 10B. Transformants generally exhibited higher protein content than wildtype.

FIG. 11 shows an induction effect by overexpressing OsRHC1 on the expression of the three rice defense marker genes. In general, the degree of induction of the three defense marker genes is positively correlated with the level of OsRHC1 expression. For instance, the two transgenic lines R8 and R12 which exhibited higher level of OsRHC1 also induced the expression of the three defense marker genes to a larger extent (comparing FIGS. 10 and 11).

EXAMPLE 9

OsRHC1 is an E3 Ubiquitin Ligase

This example demonstrates that OsRHC1 is capable of autoubiquitination, a property common to ubiquitin E3 ligases.

A partial fragment of OsRHC1 (OsRHC1p) lacking transmembrane domain located at the N-terminus was prepared. Only the RING-HC domain at the C-terminus is included as the presence of the transmembrane domains makes extraction from *E. coli* cells difficult.

The appropriate C-terminal portion of the encoding sequence was amplified with primer set HMOL5743 (5'-CCGGAATTCGTTGTTCTACTATTACGAAATGG-3') (SEQ ID NO:36) and HMOL2625 (5'-CAGGTCGACGT-TAAACATCATATACGGGCATG-3') (SEQ ID NO:37) flanking the C-terminal half containing the RING-HC domain. The PCR reaction was run with the following cycle profile: 94° C. 5 min; 30 cycles of 94° C. 30 s, 55° C. 30 s and 72° C. 1 min; followed with 72° C. extension for 5 min. The amplified product was subcloned into pGex-4T-1 vector with EcoRI and XhoI restriction sites so as to be fused with GST coding region in frame. The fusion protein was then expressed in DE3 cell with 1.5 mM IPTG induction at 30° C. for 2 hours during growth phase. GST-OsRHC1p protein was extracted by lysing the bacterial cells with 1 mg/ml lysozyme at room temperature for 1 hour, followed by 5 freeze/thaw cycles with liquid nitrogen and warm water bath. The extracted protein was purified with GST SpinTrap™ Purification Module (GH Healthcare).

The in vitro ubiquitination assay was performed in ubiquitination buffer (40 mM Tris-HCl (pH 7.5), 5 mM MgCl₂, 2 mM ATP, 2 mM dithiothreitol, 300 ng/μl ubiquitin, 25 μM MG132, 5 μl wheat germ extract (to provide E1 and E2

13

enzymes) (Promega)) plus either 400 ng GST-OsRHC1p or GST only protein. As negative control, the same reaction buffer without the addition of ATP and ubiquitin was used (modified as described by Bazirgan, O. A., et al., *J. Biol. Chem.* (2006) 281:38989-39001; Matsuda, N., et al., *J. Cell. Sci.* (2001) 114:1949-1957). The reaction mixtures were kept at room temperature for 2 hours, then subjected to 10% SDS-PAGE gel electrophoresis, and followed by Western blot analysis with anti-OsRHC1 specific antibody. (FIG. 12).

Autoubiquitination of GST-OsRHC1 was observed in the reaction including ATP and ubiquitin (+ve), but not in the reaction without ATP and ubiquitin (-ve).

These results demonstrate that, like other E3 ligases, OsRHC1 undergoes autoubiquitination.

EXAMPLE 10

Identification of an OsRHC1 Binding Partner

A protein encoded by a clone deposited in GenBank Accession No. ABA98865.1 was identified as a binding partner. This was ascertained using a yeast two hybrid protocol and verified by co-precipitation. The protein encoded by this deposited clone is expressed in *Oryza sativa* (Japonica Cultivar-Group) but it has no identified function. However, since it interacts with OsRHC1, it is presumed to modulate plant defense responses.

The yeast two hybrid protocol employed a commercial kit, the BD Matchmaker™ library construction and screening kit (Clontech K1516-1). OsRHC1 was first amplified with the oligos HMOL2624 (5'-CCGAATTCATGCCAGCCCTTCGCTTC-3') (SEQ ID NO:38) and HMOL2625 (5'-CAGGTCGACGTTAAACATCATATACGGGCATG-3') (SEQ ID NO:39), digested with the EcoRI and SalI, subcloned into pGBKT7 in reading frame and transformed into yeast strain Y187. Proteins extracted from the yeast clones transformed with pGBKT7-OsRHC1 and the control pGBKT7. Western blot analysis with anti-c-Myc epitope tag antibody confirms the presence of DNA binding domain fused OsRHC1 proteins.

Samples of RNA from several rice lines (each containing one of the following R genes: Xa2, Xa12, Xa14, Pita, Pib, and Pik) inoculated with the corresponding incompatible pathogens (T2 for Xa2; P1 for Xa12; LN44 for Xa14; Ken54-04 for Pita, Pib and Pik) for 4 days were used as starting materials to construct an AD domain fusion yeast library in the yeast strain

14

AH109 according to the manufacturer's manual. Two rounds of library screening were performed by mating between pGBKT7-OsRHC1 transformed Y187 and the AH109 yeast library. Yeast diploid mating products were selected on SD minus Trp, Leu and His (SD/-3) agar plates and incubated at 30° C. for 4 days. Only colonies grown to 2-3 mm diameter were further streaked onto SD minus Trp, Leu, His and Ade (SD/-4) agar plates. Selected clones were tested by colony-lift filter assay for lacZ reporter gene activity (*Yeast Protocols Handbook*, Clontech PT3024-1). The partial clone that encoded expressed protein (accession number: ABA98865.1) (labeled as HML1797) produced a positive result. Retransformation of pGBKT7-OsRHC1 and pGADT7-HML1797 into AH109 confirmed this was not due to mutation.

To verify the result of yeast-2-hybrid experiments, co-immunoprecipitation assays were conducted. The full-length coding region of ABA98865.1 was amplified with primers HMOL5311 (5'-AACCCGGGATGGCCGTGGGGTCA-GAG-3') (SEQ ID NO:40) and HMOL5312 (5'-TTC-CCGGGTCAAAAATAAAAACAAATAAAAAAACAC-3') (SEQ ID NO:41), digested with SmaI and subcloned into SmaI linearized pGADT7-Rec vector to generate a fusion protein with an in-frame HA tag (HA-ABA98865.1); this was designated HML1846. This construct was transcribed and translated in vitro by RiboMAX RiboMAX™ large scale RNA production systems-T7 (Promega), wheat germ extract (Promega) and Transcend™ biotin-lysyl-tRNA system (Promega) in combination, respectively.

Total protein was extracted from a rice line overexpressing OsRHC1 (modified from Boyes, D. C., et al., *Proc. Natl. Acad. Sci. USA* (1998) 95:15849-15854; Greve, K., et al., *Biochem. J.* (2003) 371:97-108. Samples from rice containing 100 µg protein were mixed with 40 µl HA tag fused protein above in a co-immunoprecipitation buffer containing 50 mM Tris/HCl (pH 7.5), 250 mM NaCl, 2 mM MgCl₂, 0.5 mM CaCl₂, 10% (v/v) glycerol, 1.5% (v/v) Triton® X-100, 1 mM PMSF, 2 mg/L leupeptin (modified from Boyes, et al., 1998, supra; Greve, et al., 2003, supra), using the BD Matchmaker™ Co-IP Kit (Clontech 630449). Anti-HA epitope tagged antibody was employed for pulling down the protein complexes. Protein signal was detected by anti-OsRHC1 antibody.

Western blot showed that OsRHC1 was pulled down by HA tag fused ABA98865.1, but no protein was detected on Western blot when the rice protein extract was treated with unrelated protein fused with HA tag.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 52

<210> SEQ ID NO 1
 <211> LENGTH: 107
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic construct
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (1)...(107)
 <223> OTHER INFORMATION: RING zing finger HC subclass consensus
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (2)...(3)
 <223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (5)...(43)

-continued

```

<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
up to 30 may be absent
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (45)...(47)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
up to 2 may be absent
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (49)...(51)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
1 may be absent
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (53)...(54)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (56)...(103)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
up to 44 may be absent
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (105)...(106)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

```

```

<400> SEQUENCE: 1

```

```

Cys Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
1           5           10          15

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
20          25          30

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa His
35          40          45

Xaa Xaa Xaa Cys Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
50          55          60

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
65          70          75          80

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
85          90          95

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
100         105

```

```

<210> SEQ ID NO 2
<211> LENGTH: 107
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic construct
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (1)...(107)
<223> OTHER INFORMATION: RING zing finger H2 subclass consensus
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (2)...(3)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (5)...(43)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
up to 30 may be absent
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (45)...(47)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
up to 2 may be absent
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (49)...(51)
<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid,
1 may be absent
<220> FEATURE:

```


-continued

<400> SEQUENCE: 6

cctcactttt gtctccac

19

<210> SEQ ID NO 7

<211> LENGTH: 18

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 7

cgacattgca caaccaac

18

<210> SEQ ID NO 8

<211> LENGTH: 35

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct

<220> FEATURE:

<221> NAME/KEY: Misc_feature

<222> LOCATION: (1)...(35)

<223> OTHER INFORMATION: RING-HC domain consensus

<220> FEATURE:

<221> NAME/KEY: Misc_feature

<222> LOCATION: (1)...(35)

<223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

<400> SEQUENCE: 8

Cys Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys
1 5 10 15Xaa His Xaa Xaa Xaa Cys Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Cys
20 25 30Xaa Xaa Cys
35

<210> SEQ ID NO 9

<211> LENGTH: 17

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic construct

<400> SEQUENCE: 9

Asn Cys Gly Tyr Pro Pro Glu Val Val Arg Lys Met Pro Lys Arg Asp
1 5 10 15

Cys

<210> SEQ ID NO 10

<211> LENGTH: 22

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 10

aaagaagagc aagcccggtt at

22

<210> SEQ ID NO 11

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 11

-continued

gcctccatac ctcttctgca a 21

<210> SEQ ID NO 12
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 12

cttcatagga atggaagctg cgggta 26

<210> SEQ ID NO 13
<211> LENGTH: 25
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 13

gaccaccttg atcttcatgc tgcta 25

<210> SEQ ID NO 14
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 14

tcaagatagc ccacaagatt atc 23

<210> SEQ ID NO 15
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 15

cttctcggtc acataattcc cac 23

<210> SEQ ID NO 16
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 16

accaccactg atacgtctcc tc 22

<210> SEQ ID NO 17
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 17

aacttcatac ttagactgtc gatc 24

<210> SEQ ID NO 18
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:

-continued

```

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 18

cccttatctt cgctgctctt gt                22

<210> SEQ ID NO 19
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 19

ccctgaccat gtcccacttg                20

<210> SEQ ID NO 20
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 20

agcactgcaa gttagggtgt ga                22

<210> SEQ ID NO 21
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 21

acattgttcc gacgctccat                20

<210> SEQ ID NO 22
<211> LENGTH: 19
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 22

gaaggtgctg agttgattg                19

<210> SEQ ID NO 23
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 23

ggacttgacg ttgtttgg                18

<210> SEQ ID NO 24
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Forward primer

<400> SEQUENCE: 24

ctgatgcata tacatgatgg                20

<210> SEQ ID NO 25

```

-continued

<211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Reverse primer

 <400> SEQUENCE: 25

 acattgcaca accaacaatgt ac 22

<210> SEQ ID NO 26
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Forward primer

 <400> SEQUENCE: 26

 aaagaagagc aagcccggtt at 22

<210> SEQ ID NO 27
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Reverse primer

 <400> SEQUENCE: 27

 gcctccatac ctcttctgca a 21

<210> SEQ ID NO 28
 <211> LENGTH: 26
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Forward primer

 <400> SEQUENCE: 28

 cggacagagg ccttactaag ttattt 26

<210> SEQ ID NO 29
 <211> LENGTH: 28
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Reverse primer

 <400> SEQUENCE: 29

 gacctgttta cattttcacg tctttatt 28

<210> SEQ ID NO 30
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Forward primer

 <400> SEQUENCE: 30

 gaggcaacgg acaccactaa g 21

<210> SEQ ID NO 31
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Reverse primer

 <400> SEQUENCE: 31

-continued

tgtaaagcag agagagaggc tcatt 25

<210> SEQ ID NO 32
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Forward primer

<400> SEQUENCE: 32

aagctcaagt cacactcgac 20

<210> SEQ ID NO 33
<211> LENGTH: 19
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Reverse primer

<400> SEQUENCE: 33

gatgtccttc tccttctcc 19

<210> SEQ ID NO 34
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Forward primer

<400> SEQUENCE: 34

cttcatagga atggaagctg cgggta 26

<210> SEQ ID NO 35
<211> LENGTH: 25
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Reverse primer

<400> SEQUENCE: 35

gaccaccttg atcttcatgc tgcta 25

<210> SEQ ID NO 36
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 36

ccggaattcg ttgttctact attacgaaat gg 32

<210> SEQ ID NO 37
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 37

caggtcgacg ttaaacaatca tatacgggca tg 32

<210> SEQ ID NO 38
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:

-continued

```

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 38

ccgaattcat gccagccct tcgcttc                27

<210> SEQ ID NO 39
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 39

caggtcgacg ttaaacatca tatacgggca tg        32

<210> SEQ ID NO 40
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 40

aaccgggat gcccgtaggg tcagag                26

<210> SEQ ID NO 41
<211> LENGTH: 35
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 41

ttccggggtc aaaataaaaa caaataaaaa aacac    35

<210> SEQ ID NO 42
<211> LENGTH: 1230
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic construct
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (1)...(1230)
<223> OTHER INFORMATION: RING zinc finger protein gene (OsRHC1)
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)...(1230)

<400> SEQUENCE: 42

atg cca gcc cct tcg ctt cct cat ggc cgt cat tgg gct cct tgc cat    48
Met Pro Ala Pro Ser Leu Pro His Gly Arg His Trp Ala Pro Cys His
1          5          10          15

tca att gtt gca gcg ccg ttg ctt att gcg ttt gag ctg ctg ctt tgc    96
Ser Ile Val Ala Ala Pro Leu Leu Ile Ala Phe Glu Leu Leu Leu Cys
20          25          30

ata tat ctc gaa agt ttg aga gtt aaa agt aag ccg act gtt gat ttg    144
Ile Tyr Leu Glu Ser Leu Arg Val Lys Ser Lys Pro Thr Val Asp Leu
35          40          45

aag att gta ttc ctt cct ctt ctg gcc ttt gaa gtg att att ctt gtt    192
Lys Ile Val Phe Leu Pro Leu Leu Ala Phe Glu Val Ile Ile Leu Val
50          55          60

gac aat ttc aga atg tgt aga gct tta atg cca gga gat gaa gaa agt    240
Asp Asn Phe Arg Met Cys Arg Ala Leu Met Pro Gly Asp Glu Glu Ser
65          70          75          80

atg agc gat gaa gct att tgg gag aca ctt cct cac ttt tgg gtt gca    288

```


-continued

Glu Glu Arg Met Pro Val Tyr Asp Val
405

<210> SEQ ID NO 43
<211> LENGTH: 409
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic construct
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (1)...(409)
<223> OTHER INFORMATION: RING zinc finger protein gene (OsRHCl)

<400> SEQUENCE: 43

Met Pro Ala Pro Ser Leu Pro His Gly Arg His Trp Ala Pro Cys His
1 5 10 15
Ser Ile Val Ala Ala Pro Leu Leu Ile Ala Phe Glu Leu Leu Leu Cys
20 25 30
Ile Tyr Leu Glu Ser Leu Arg Val Lys Ser Lys Pro Thr Val Asp Leu
35 40 45
Lys Ile Val Phe Leu Pro Leu Leu Ala Phe Glu Val Ile Ile Leu Val
50 55 60
Asp Asn Phe Arg Met Cys Arg Ala Leu Met Pro Gly Asp Glu Glu Ser
65 70 75 80
Met Ser Asp Glu Ala Ile Trp Glu Thr Leu Pro His Phe Trp Val Ala
85 90 95
Ile Ser Met Val Phe Leu Ile Ala Ala Thr Thr Phe Thr Leu Leu Lys
100 105 110
Leu Ser Gly Asp Val Gly Ala Leu Gly Trp Trp Asp Leu Phe Ile Asn
115 120 125
Tyr Gly Ile Ala Glu Cys Phe Ala Phe Leu Val Cys Thr Arg Trp Phe
130 135 140
Asn Pro Met Ile His Lys Ser Pro Asn Pro Gly Glu Ala Ser Ser Ser
145 150 155 160
Ser Ala Ala Ile Arg Tyr Arg Asp Trp Glu Ser Gly Leu Leu Leu Pro
165 170 175
Ser Leu Glu Asp His Glu Gln Glu Arg Leu Cys Gly Leu Pro Asp Ile
180 185 190
Gly Gly His Val Met Lys Ile Pro Leu Val Ile Phe Gln Val Leu Leu
195 200 205
Cys Met Arg Leu Glu Gly Thr Pro Pro Ser Ala Gln Tyr Ile Pro Ile
210 215 220
Phe Ala Leu Phe Ser Pro Leu Phe Ile Leu Gln Gly Ala Gly Val Leu
225 230 235 240
Phe Ser Leu Ala Arg Leu Leu Glu Lys Val Val Leu Leu Leu Arg Asn
245 250 255
Gly Pro Val Ser Pro Asn Tyr Leu Thr Ile Ser Ser Lys Val Arg Asp
260 265 270
Cys Phe Ala Phe Leu His Arg Gly Ser Arg Leu Leu Gly Trp Trp Ser
275 280 285
Ile Asp Glu Gly Ser Lys Glu Glu Gln Ala Arg Leu Phe Tyr Thr Glu
290 295 300
Ser Thr Gly Tyr Asn Thr Phe Cys Gly Tyr Pro Pro Glu Val Val Arg
305 310 315 320
Lys Met Pro Lys Arg Asp Leu Ala Glu Glu Val Trp Arg Leu Gln Ala
325 330 335

-continued

Ala Leu Gly Glu Gln Ser Glu Ile Thr Lys Cys Thr Lys Gln Glu Phe
 340 345 350

Glu Arg Leu Gln Asn Glu Lys Val Leu Cys Arg Ile Cys Tyr Glu Gly
 355 360 365

Glu Ile Cys Met Val Leu Leu Pro Cys Arg His Arg Thr Leu Cys Lys
 370 375 380

Thr Cys Ser Asp Lys Cys Lys Lys Cys Pro Ile Cys Arg Val Pro Ile
 385 390 395 400

Glu Glu Arg Met Pro Val Tyr Asp Val
 405

<210> SEQ ID NO 44
 <211> LENGTH: 467
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic construct
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (1)...(467)
 <223> OTHER INFORMATION: Annotated protein NP-564052

<400> SEQUENCE: 44

Met Ser Cys Arg Arg Val Leu Lys Ser Ile Gln Ala Leu Ala Ala His
 1 5 10 15

Ser Leu Leu Phe Cys Phe Thr Leu Leu Val Leu Lys Leu Asp His
 20 25 30

Thr Val Ser Ser Ser Trp Trp Met Val Phe Phe Pro Leu Trp Ala Phe
 35 40 45

His Ala Val Val Ala Arg Gly Arg Phe Ser Leu Pro Ala Pro Val Ala
 50 55 60

Pro Arg Asn Arg His Trp Ala Pro Cys His Ala Val Val Ala Thr Pro
 65 70 75 80

Leu Leu Val Ala Phe Glu Leu Leu Leu Cys Ile Tyr Leu Glu Ser Ser
 85 90 95

Tyr Ala Arg Trp Pro Pro Ala Val Ser Leu Lys Ile Ala Phe Leu Pro
 100 105 110

Leu Leu Ala Phe Glu Leu Thr Ile Leu Val Asp Asn Leu Arg Met Cys
 115 120 125

Arg Ala Leu Met Pro Gly Asp Asp Ser Ile Thr Asp Asp Ala Ile
 130 135 140

Trp Glu Ala Leu Pro His Phe Trp Val Ala Ile Ser Met Val Phe Thr
 145 150 155 160

Leu Ala Ala Thr Phe Phe Thr Leu Leu Lys Leu Ser Gly Asp Val Val
 165 170 175

Ala Leu Gly Trp Trp Asp Leu Phe Ile Asn Phe Gly Ile Ala Glu Cys
 180 185 190

Phe Ala Phe Leu Val Cys Thr Lys Trp Ser Asn Pro Val Ile His Arg
 195 200 205

Ser Ser Arg Ala Arg Glu Thr Gly Ser Ser Ser Thr Ser Ile Arg Tyr
 210 215 220

Leu Asp Trp Asn Ser Gly Leu Val Val Ala Pro Glu Glu Asp Arg His
 225 230 235 240

Gln Asp Arg Trp Cys Gly Leu Gln Asp Ile Gly Gly His Met Leu Lys
 245 250 255

Ile Pro Val Ile Leu Phe Gln Val Val Leu Cys Met Tyr Leu Glu Gly
 260 265 270

-continued

Trp Glu Ala Leu Pro His Phe Trp Val Ala Ile Ser Met Val Phe Phe
 145 150 155 160
 Leu Ala Ala Thr Val Phe Thr Leu Leu Lys Leu Ser Gly Asp Val Ala
 165 170 175
 Ala Leu Gly Trp Trp Asp Leu Phe Ile Asn Phe Gly Ile Ala Glu Cys
 180 185 190
 Phe Ala Phe Leu Val Cys Thr Lys Trp Ser Asn Pro Val Ile His Arg
 195 200 205
 Ser Ser Arg Asp Arg Glu Thr Gly Ser Ser Ser Thr Asn Ile Arg Tyr
 210 215 220
 Leu Asp Trp Asn Ser Gly Leu Gly Val Phe Ser Glu Asp Asp Arg Asn
 225 230 235 240
 Gln Asp Thr Cys Gly Leu Gln Asp Ile Gly Gly His Ile Met Lys Ile
 245 250 255
 Pro Leu Ile Val Phe Gln Val Val Leu Cys Met His Leu Glu Gly Thr
 260 265 270
 Pro Glu Ala Ala Lys Ser Ile Ser Val Pro Val Leu Phe Ser Pro Leu
 275 280 285
 Phe Leu Leu Gln Gly Val Gly Val Leu Phe Ala Ala Ser Lys Leu Ile
 290 295 300
 Glu Lys Val Val Leu Leu Leu Arg Gly Glu Asp Asp Thr Gly Leu Tyr
 305 310 315 320
 Phe Arg Phe Leu Ser Arg Ala His Asp Cys Leu Gly Phe Leu His His
 325 330 335
 Gly Ser Arg Leu Leu Gly Trp Trp Ser Ile Asp Glu Gly Ser Arg Glu
 340 345 350
 Glu Glu Ala Arg Leu Tyr Phe Asp Gln Glu Ser Gly Tyr Asn Thr Phe
 355 360 365
 Cys Gly His Pro Pro Glu Ile Val Lys Lys Met Pro Lys Lys Glu Leu
 370 375 380
 Ala Glu Glu Val Trp Arg Leu Gln Ala Ala Leu Gly Glu Gln Thr Glu
 385 390 395 400
 Ile Thr Lys Phe Ser Gln Gln Glu Tyr Glu Arg Leu Gln Asn Glu Lys
 405 410 415
 Val Leu Cys Arg Val Cys Phe Glu Arg Glu Ile Ser Val Val Leu Leu
 420 425 430
 Pro Cys Arg His Arg Val Leu Cys Arg Asn Cys Ser Asp Lys Cys Lys
 435 440 445
 Lys Cys Pro Phe Cys Arg Ile Thr Ile Glu Glu Arg Leu Pro Val Tyr
 450 455 460
 Asp Val
 465

<210> SEQ ID NO 46
 <211> LENGTH: 467
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic construct
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (1)...(467)
 <223> OTHER INFORMATION: Annotated protein AAW81737
 <400> SEQUENCE: 46

Met Ser Cys Arg Arg Val Leu Lys Ser Ile Gln Ala Leu Ala Ala His
 1 5 10 15

-continued

Ser Leu Leu Phe Ser Phe Thr Leu Phe Leu Val Phe Lys Leu Asp His
 20 25 30
 Thr Leu Ser Cys Ser Trp Trp Met Val Phe Phe Pro Leu Trp Ala Phe
 35 40 45
 His Ala Val Val Ala Arg Gly Arg Phe Ser Leu Pro Ala Pro Ile Ala
 50 55 60
 Pro Arg Asn Arg His Trp Ala Pro Cys His Ala Val Val Ala Thr Pro
 65 70 75 80
 Leu Leu Val Ser Phe Glu Leu Leu Leu Cys Ile Tyr Leu Glu Ser Ser
 85 90 95
 Tyr Ala Ser Trp Pro Pro Ala Val Ser Leu Arg Ile Ala Ser Leu Pro
 100 105 110
 Leu Leu Ala Phe Glu Val Thr Ile Leu Ile Asp Asn Leu Arg Met Cys
 115 120 125
 Arg Ala Leu Met Pro Gly Asp Asp Asp Ser Ile Asn Asp Glu Ala Ile
 130 135 140
 Trp Glu Ala Leu Pro His Phe Trp Val Ala Ile Ser Met Val Phe Thr
 145 150 155 160
 Leu Ala Ala Thr Phe Phe Ala Leu Leu Lys Leu Thr Gly Asp Val Ala
 165 170 175
 Ala Leu Ser Trp Trp Asp Leu Phe Ile Asn Val Gly Ile Ala Glu Cys
 180 185 190
 Phe Ala Phe Leu Val Cys Thr Lys Trp Ser Asn Pro Val Ile His Arg
 195 200 205
 Ser Ser Arg Pro Arg Glu Thr Gly Ser Ser Ser Thr Pro Val Arg Tyr
 210 215 220
 Leu Asp Trp Asn Ser Gly Leu Val Val Thr Pro Glu Gln Asp Asn His
 225 230 235 240
 Gln Asp Arg Tyr Cys Gly Leu Gln Asp Ile Gly Gly His Leu Leu Lys
 245 250 255
 Ile Pro Val Ile Val Phe Gln Val Val Leu Cys Met His Leu Glu Gly
 260 265 270
 Thr Pro Glu Arg Ala Lys Asp Ile Ser Ile Pro Val Leu Phe Ser Pro
 275 280 285
 Ile Phe Leu Leu Gln Gly Leu Gly Val Leu Phe Ala Thr Ser Lys Leu
 290 295 300
 Ile Glu Lys Ile Val Asp Leu Leu Gln Gly Glu Ala Gly Thr Gly Leu
 305 310 315 320
 Tyr Phe Arg Val Ser Ser Arg Ala His Asp Cys Leu Gly Phe Leu His
 325 330 335
 His Gly Ser Arg Leu Leu Gly Trp Trp Ser Ile Asp Glu Gly Ser Arg
 340 345 350
 Glu Glu Gln Ala Arg Leu Tyr Phe Asp Gln Glu Ser Gly Tyr Asn Thr
 355 360 365
 Phe Ser Gly His Pro Pro Glu Ile Val Lys Lys Met Pro Lys Glu Asp
 370 375 380
 Leu Ala Glu Glu Val Trp Arg Leu Gln Ala Ala Leu Gly Glu Gln Thr
 385 390 395 400
 Glu Ile Thr Lys Phe Ser Gln Gln Glu Tyr Glu Arg Leu Gln Asn Glu
 405 410 415
 Lys Val Leu Cys Arg Val Cys Phe Glu Lys Glu Ile Ser Leu Val Leu
 420 425 430
 Leu Pro Cys Arg His Arg Val Leu Cys Arg Ile Cys Ser Asp Lys Cys
 435 440 445

-continued

Thr Lys Cys Pro Ile Cys Arg Val Ala Ile Glu Glu Arg Leu Leu Val
 450 455 460

Tyr Asp Val
 465

<210> SEQ ID NO 47
 <211> LENGTH: 466
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic construct
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (1)...(466)
 <223> OTHER INFORMATION: Annotated protein BAE71207

<400> SEQUENCE: 47

Met Ser Trp Ser Arg Val Leu Lys Ser Ala Gln Ala Phe Ala Ala His
 1 5 10 15
 Thr Phe Leu Leu Cys Phe Thr Leu Leu Leu Leu Leu Lys Leu Asp His
 20 25 30
 Gln Ile Ser Ser Ser Trp Trp Ile Ile Phe Ser Pro Leu Trp Met Phe
 35 40 45
 His Gly Val Val Ala Arg Gly Arg Phe Ser Leu Pro Ala Pro Ser Ala
 50 55 60
 Pro Arg Asn Arg His Trp Ala Pro Cys His Ala Val Val Ala Met Pro
 65 70 75 80
 Leu Leu Ile Ala Phe Glu Leu Leu Leu Cys Ile Tyr Leu Glu Ser Leu
 85 90 95
 Tyr Val Arg Gly Phe Pro Ala Val Asp Leu Lys Ile Val Phe Leu Pro
 100 105 110
 Leu Leu Thr Phe Glu Val Ile Ile Leu Ile Asp Asn Phe Arg Met Cys
 115 120 125
 Lys Ala Leu Met Pro Gly Asp Glu Glu Arg Met Ser Asp Glu Ala Ile
 130 135 140
 Trp Glu Thr Leu Pro His Phe Trp Val Ala Ile Ser Met Val Phe Phe
 145 150 155 160
 Val Ala Ala Thr Val Phe Thr Leu Leu Lys Leu Ser Gly Ser Val Ala
 165 170 175
 Ser Leu Gly Trp Trp Asp Leu Phe Ile Asn Phe Thr Ile Ala Glu Cys
 180 185 190
 Phe Ala Phe Leu Val Cys Thr Lys Trp Ser Asn Pro Val Ile His Arg
 195 200 205
 Ser Ser Arg Glu Pro Ser Ser Ser Ser Ser Thr Thr Ile Arg Tyr Leu
 210 215 220
 Asp Trp Asn Asn Gly Leu Leu Val Ser Ser Glu Glu Asp Gln Arg Gln
 225 230 235 240
 Ala Arg Ile Cys Thr Leu Gln Asp Ile Gly Gly His Phe Met Lys Val
 245 250 255
 Pro Ile Ile Val Phe Gln Val Leu Leu Cys Met His Leu Glu Gly Thr
 260 265 270
 Pro Ala Phe Ala Ala Gln Leu Pro Leu Ala Val Leu Phe Ser Pro Leu
 275 280 285
 Phe Val Leu Gln Gly Val Gly Val Ile Leu Ser Ala Ser Lys Phe Val
 290 295 300
 Glu Lys Leu Val Leu Leu Leu Arg Ser Gly Ala Gly Gly Gly Leu Tyr

-continued

```

305              310              315              320
Phe Arg Val Ser Ser Ile Ala His Asp Cys Leu Gly Phe Leu His His
              325              330              335
Gly Ser Arg Leu Leu Gly Trp Trp Ser Ile Asp Glu Gly Ser Arg Glu
              340              345              350
Glu Gln Ala Arg Leu Tyr His Glu Gly Ala Ser Gly Tyr Asn Thr Phe
              355              360              365
Ser Gly Tyr Pro Pro Glu Ile Val Lys Lys Met Pro Lys Arg Asp Leu
              370              375              380
Ala Glu Glu Val Trp Arg Leu Gln Ala Ala Leu Gly Glu Gln Thr Glu
385              390              395              400
Ile Thr Lys Tyr Ser Gln Gln Glu Tyr Glu Arg Leu Lys Asn Glu Lys
              405              410              415
Val Leu Cys Arg Ile Cys Phe Glu Gly Glu Ile Ser Val Val Leu Leu
              420              425              430
Pro Cys Arg His Arg Val Leu Cys Ser Leu Cys Ser Glu Lys Cys Lys
              435              440              445
Met Cys Pro Ile Cys Arg Asn Tyr Ile Ala Glu Arg Leu Pro Val Tyr
              450              455              460

Asp Val
465

```

```

<210> SEQ ID NO 48
<211> LENGTH: 468
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic construct
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (1)...(468)
<223> OTHER INFORMATION: Annotated protein NP_564945

<400> SEQUENCE: 48

```

```

Met Leu Val Gln Arg Arg Val Met Ser Trp Arg Arg Val Trp Lys Ser
 1              5              10              15
Phe Gln Ala Ala Ser Ala His Cys Leu Leu Phe Ser Phe Thr Leu Leu
              20              25              30
Leu Ala Leu Lys Leu Asp His Val Val Ser His Ser Trp Trp Phe Val
              35              40              45
Phe Ala Pro Leu Trp Leu Phe His Ala Val Ile Ala Arg Gly Arg Phe
 50              55              60
Ser Leu Pro Ala Pro Ser Met Pro His Asp Arg His Trp Ala Pro Phe
65              70              75              80
His Ser Val Met Ala Thr Pro Leu Leu Val Ala Phe Glu Ile Leu Leu
              85              90              95
Cys Val His Leu Glu Asp Lys Tyr Val Val Asp Leu Lys Ile Val Phe
              100              105              110
Leu Pro Leu Leu Ala Phe Glu Val Ala Ile Leu Ile Asp Asn Val Arg
              115              120              125
Met Cys Arg Thr Leu Met Pro Gly Asp Glu Glu Thr Met Ser Asp Glu
130              135              140
Ala Ile Trp Glu Thr Leu Pro His Phe Trp Val Ser Ile Ser Met Val
145              150              155              160
Phe Phe Ile Ala Ala Thr Thr Phe Thr Leu Leu Lys Leu Cys Gly Asp
              165              170              175
Val Ala Ala Leu Gly Trp Trp Asp Leu Phe Ile Asn Phe Gly Ile Ala

```

-continued

```

      180              185              190
Glu Cys Phe Ala Phe Leu Val Cys Thr Lys Trp Ser Asn Gln Ser Ile
   195                200                205
His Arg Tyr Ser His Ile Pro Glu Pro Ser Ser Ser Ser Met Val Val
   210                215                220
Arg Tyr Leu Asp Trp Asn Arg Gly Leu Val Val Thr Ala Asp Asp Glu
   225                230                235
His Gln Gln Ser Asn Arg Ile Cys Gly Leu Gln Asp Ile Gly Gly His
   245                250                255
Val Met Lys Ile Pro Phe Val Thr Phe Gln Ile Ile Leu Phe Met Arg
   260                265                270
Leu Glu Gly Thr Pro Ala Ser Ala Lys Asn Ile Pro Ile Leu Val Leu
   275                280                285
Phe Val Pro Leu Phe Leu Leu Gln Gly Ala Gly Val Leu Phe Ala Met
   290                295                300
Tyr Arg Leu Val Glu Lys Ser Val Leu Leu Ile Asn Ser Gly Ser Gly
   305                310                315
Ser Tyr Gly Arg Tyr Phe Thr Ala Thr Ser Ser Ala Arg Glu Phe Leu
   325                330                335
Gly Phe Phe Gln His Gly Ala Arg Leu Leu Gly Trp Trp Ser Ile Asp
   340                345                350
Glu Gly Ser Arg Glu Glu Gln Ala Arg Leu Tyr Ser Gly Glu Ala Thr
   355                360                365
Gly Tyr Asn Thr Phe Ser Pro Glu Val Val Lys Lys Met Pro Lys Ser
   370                375                380
Asp Leu Val Glu Glu Ile Trp Arg Leu Gln Ala Ala Leu Ser Glu Gln
   385                390                395
Thr Asp Ile Thr Ser Tyr Ser Gln Gln Glu Tyr Glu Arg Leu Gln Asn
   405                410                415
Glu Lys Ile Leu Cys Arg Val Cys Phe Glu Asp Pro Ile Asn Val Val
   420                425                430
Leu Leu Pro Cys Arg His His Val Leu Cys Ser Thr Cys Cys Glu Lys
   435                440                445
Cys Lys Lys Cys Pro Ile Cys Arg Val Leu Ile Glu Glu Arg Met Pro
   450                455                460
Val Tyr Asp Val
465

```

```

<210> SEQ ID NO 49
<211> LENGTH: 497
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic construct
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (1)...(497)
<223> OTHER INFORMATION: Annotated protein ABE90658

```

```

<400> SEQUENCE: 49

```

```

Met Leu Val Arg Arg Arg Val Met Ser Trp Arg Arg Val Phe Lys Ser
 1              5              10              15
Leu Gln Ala Met Leu Ala His Ala Phe Leu Phe Ser Phe Ser Leu Leu
 20             25             30
Leu Val Leu Lys Leu Asp Arg Phe Phe Leu Phe Ser Trp Trp Thr Val
 35             40             45
Phe Phe Pro Leu Trp Leu Phe His Val Val Ile Ala Arg Gly Arg Phe

```

-continued

50					55					60						
Ser	Leu	Pro	Ala	Pro	Ser	Met	Pro	His	Gly	Arg	Gln	Trp	Ala	Pro	Cys	
65					70					75					80	
His	Ser	Val	Ile	Ala	Thr	Pro	Leu	Leu	Val	Ala	Phe	Glu	Leu	Leu	Leu	
			85						90					95		
Cys	Ile	His	Leu	Gly	Ser	Ser	Tyr	Val	Val	Asn	Leu	Lys	Ile	Val	Phe	
			100					105					110			
Ile	Pro	Leu	Ile	Ala	Phe	Glu	Leu	Ala	Ile	Leu	Ile	Asp	Asn	Ile	Arg	
		115					120					125				
Met	Cys	Arg	Ala	Leu	Met	Pro	Gly	Asp	Glu	Glu	Asn	Met	Thr	Asp	Glu	
		130					135					140				
Ala	Val	Trp	Glu	Thr	Leu	Pro	His	Phe	Trp	Ile	Ser	Ile	Ser	Met	Val	
		145					150					155				
Phe	Phe	Val	Ala	Ala	Thr	Val	Phe	Thr	Leu	Lys	Ile	Cys	Gly	Asp		
			165						170				175			
Val	Ala	Ala	Leu	Gly	Trp	Trp	Asp	Leu	Phe	Ile	Asn	Tyr	Gly	Tyr	Asn	
			180					185					190			
Gln	Tyr	Leu	Leu	Val	Asp	Cys	Phe	Lys	His	Phe	Ile	Leu	Ile	Leu	Tyr	
		195					200					205				
Phe	Phe	His	His	Lys	Leu	Ile	Leu	Ser	Phe	Cys	Ser	Ile	Ala	Gln	Cys	
		210					215					220				
Phe	Ala	Phe	Leu	Val	Cys	Thr	Lys	Trp	His	Asn	Pro	Thr	Ile	His	Gly	
		225					230					235			240	
Asn	Gly	His	Ile	Thr	Glu	Pro	Cys	Ser	Ser	Ser	Asn	Thr	Val	Arg	Tyr	
			245						250					255		
Leu	Glu	Trp	Ser	Arg	Glu	Gly	Ile	Val	Ile	Ser	Thr	Glu	Glu	Asp	Glu	
			260				265						270			
Gln	Gln	Asn	Val	Phe	Cys	Ser	Leu	Gln	Asp	Ile	Gly	Gly	His	Ile	Met	
		275					280					285				
Lys	Ile	Pro	Phe	Ile	Ala	Phe	Gln	Ile	Leu	Leu	Phe	Met	His	Leu	Glu	
		290					295					300				
Gly	Thr	Pro	Ser	Gly	Ala	Lys	Asp	Ile	Pro	Ile	Trp	Val	Ile	Phe	Ser	
		305					310					315			320	
Pro	Leu	Leu	Leu	Leu	Gln	Gly	Ala	Gly	Val	Leu	Phe	Ala	Ala	Tyr	Arg	
				325					330					335		
Leu	Ile	Glu	Lys	Ile	Ile	Leu	Leu	Leu	Tyr	Asn	Gly	Asp	Ile	Pro	Arg	
			340					345					350			
Ser	Tyr	Ser	Ser	Ile	Ser	Ser	Lys	Ser	Arg	Asp	Cys	Phe	Gly	Phe	Phe	
		355					360					365				
Asn	His	Gly	Ser	Arg	Leu	Leu	Gly	Trp	Trp	Ser	Ile	Asp	Glu	Gly	Ser	
		370					375					380				
Arg	Glu	Glu	Glu	Ala	Arg	Leu	Phe	Cys	Ala	Gly	Ser	Ser	Gly	Tyr	Asn	
				385			390					395			400	
Thr	Phe	Ser	Pro	Asp	Thr	Val	Lys	Lys	Met	Pro	Arg	Gly	Glu	Leu	Val	
				405					410					415		
Glu	Glu	Ile	Trp	Arg	Leu	Gln	Ala	Ala	Leu	Gly	Glu	Gln	Thr	Glu	Val	
			420					425					430			
Thr	Lys	Tyr	Ser	Gln	Glu	Glu	Tyr	Glu	Arg	Leu	Gln	Asn	Glu	Lys	Ile	
		435					440					445				
Leu	Cys	Arg	Val	Cys	Phe	Glu	Glu	Gln	Ile	Asn	Val	Val	Leu	Leu	Pro	
		450					455					460				
Cys	Lys	His	His	Val	Leu	Cys	Ser	Thr	Cys	Cys	Glu	Lys	Cys	Lys	Lys	
				465			470					475			480	

-continued

Cys Pro Ile Cys Arg Gly Thr Ile Glu Glu Arg Met Pro Ile Tyr Asp
485 490 495

Val

<210> SEQ ID NO 50
 <211> LENGTH: 498
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic construct
 <220> FEATURE:
 <221> NAME/KEY: Misc_feature
 <222> LOCATION: (1)...(498)
 <223> OTHER INFORMATION: Annotated protein AAF25982

<400> SEQUENCE: 50

Met Val Phe Phe Pro Leu Trp Ala Phe His Ala Val Val Ala Arg Gly
1 5 10 15
 Arg Phe Ser Leu Pro Ala Pro Val Ala Pro Arg Asn Arg His Trp Ala
20 25 30
 Pro Cys His Ala Val Val Ala Thr Pro Leu Leu Val Ala Phe Glu Leu
35 40 45
 Leu Leu Cys Ile Tyr Leu Glu Ser Ser Tyr Ala Arg Trp Pro Pro Ala
50 55 60
 Val Ser Leu Lys Ile Ala Phe Leu Pro Leu Leu Ala Phe Glu Leu Thr
65 70 75 80
 Ile Leu Val Asp Asn Leu Arg Met Cys Arg Ala Leu Met Pro Gly Asp
85 90 95
 Asp Asp Ser Ile Thr Asp Asp Ala Ile Trp Glu Ala Leu Pro Val Ser
100 105 110
 Pro Leu Leu Leu His Lys Ile Phe Glu Gly Leu Ser Leu Arg Leu Gly
115 120 125
 Lys Ile Asn Leu Leu Asn Met Asn Glu Asn Leu Ser Leu Ile Phe Gln
130 135 140
 Leu His Asn Ser Gly Leu Arg Arg Glu Lys Thr Leu Thr Asn His Phe
145 150 155 160
 Trp Val Ala Ile Ser Met Val Phe Thr Leu Ala Ala Thr Phe Phe Thr
165 170 175
 Leu Leu Lys Leu Ser Val Phe Glu Lys Tyr Leu Pro Phe Leu Trp Leu
180 185 190
 Leu Val Lys Asn Met Lys Val Ile Tyr Met Lys Cys Ser Ala Cys Arg
195 200 205
 Ile Ala Glu Cys Phe Ala Phe Leu Val Cys Thr Lys Trp Ser Asn Pro
210 215 220
 Val Ile His Arg Ser Ser Arg Ala Arg Glu Thr Gly Ser Ser Ser Thr
225 230 235 240
 Ser Ile Arg Tyr Leu Asp Trp Asn Ser Gly Leu Val Val Ala Pro Glu
245 250 255
 Glu Asp Arg His Gln Asp Arg Trp Cys Gly Leu Gln Asp Ile Gly Gly
260 265 270
 His Met Leu Lys Ile Pro Val Ile Leu Phe Gln Val Val Leu Cys Met
275 280 285
 Tyr Leu Glu Gly Thr Pro Glu Arg Ala Lys Asp Ile Ser Ile Pro Val
290 295 300
 Leu Phe Ser Pro Leu Phe Leu Leu Gln Gly Leu Gly Val Leu Phe Ala
305 310 315 320
 Ala Ser Lys Leu Leu Glu Lys Ile Val Leu Leu Leu Arg Gly Glu Ala

-continued

	325		330		335										
Gly	Pro	Gly	Leu	Tyr	Phe	Arg	Phe	Ser	Ser	Ser	Ala	His	Asp	Cys	Leu
			340						345				350		
Gly	Phe	Leu	His	His	Gly	Ser	Arg	Leu	Leu	Gly	Trp	Trp	Ser	Ile	Asp
		355					360					365			
Glu	Gly	Ser	Arg	Glu	Glu	Gln	Ala	Arg	Leu	Tyr	Phe	Asp	Gln	Glu	Ser
	370					375					380				
Gly	Leu	Val	Trp	Arg	Leu	Gln	Ala	Ala	Leu	Gly	Glu	Gln	Thr	Glu	Ile
385					390					395					400
Thr	Lys	Phe	Ser	Gln	Glu	Tyr	Glu	Arg	Leu	Gln	Asn	Val	Tyr	Ser	
			405					410					415		
Phe	Ile	Ser	His	Asp	Val	Phe	Val	Thr	Phe	Leu	Phe	Arg	Phe	Tyr	Phe
			420					425					430		
Phe	Pro	Leu	Leu	Asn	Pro	Val	Ser	Met	Cys	Leu	Leu	Leu	Gln	Glu	Lys
		435					440						445		
Val	Leu	Cys	Arg	Val	Cys	Phe	Glu	Lys	Asp	Ile	Ser	Leu	Val	Leu	Leu
	450					455					460				
Pro	Cys	Arg	His	Arg	Val	Leu	Cys	Arg	Thr	Cys	Ala	Asp	Lys	Cys	Thr
465					470					475					480
Thr	Cys	Pro	Ile	Cys	Arg	Ile	Asp	Ile	Glu	Lys	Arg	Leu	Ser	Val	Tyr
			485						490					495	

Asp Val

<210> SEQ ID NO 51

<211> LENGTH: 354

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic construct

<220> FEATURE:

<221> NAME/KEY: Misc_feature

<222> LOCATION: (1)...(354)

<223> OTHER INFORMATION: OsRHC1 binding partner

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (1)...(354)

<400> SEQUENCE: 51

atg gcc gtg ggg tca gag cgg ctc ggc gag gag gcc gcc cgg cgg cag	48
Met Ala Val Gly Ser Glu Arg Leu Gly Glu Glu Ala Ala Arg Arg Gln	
1 5 10 15	
ctc gcc gag gca agg aag gcc aga ggc gcc tgc tgc gcg acg agg gac	96
Leu Gly Glu Ala Arg Lys Ala Arg Gly Gly Cys Ser Ala Thr Arg Asp	
20 25 30	
ggc gcc gat gat gag ggc cgg cgg cag ata aac cct ccc tcc ccg gtg	144
Gly Ala Asp Asp Glu Gly Arg Arg Gln Ile Asn Pro Pro Ser Pro Val	
35 40 45	
tgg tgg tcc cct ccc tca ctc cct ctt cct ctc aga tct gcc cgg agg	192
Trp Ser Ser Pro Pro Ser Leu Pro Leu Pro Leu Arg Ser Ala Arg Arg	
50 55 60	
ggg acg ggt gga ggc cgg cgg cct ccc ttc cct ctt tcc tct cag atc	240
Gly Thr Gly Gly Gly Arg Arg Pro Pro Phe Pro Leu Ser Ser Gln Ile	
65 70 75 80	
cgc ccg gtg ggg aga ggc cac cgg cgg cag cgg cat ggc cct ccc ctc	288
Arg Pro Val Gly Arg Gly His Arg Arg Gln Arg His Gly Pro Pro Leu	
85 90 95	
tgc agc agt aga ggg cgg cag gga gga ggc cac aga gct gtg ttt ttt	336
Cys Ser Ser Arg Gly Arg Gln Gly Gly Gly His Arg Ala Val Phe Phe	
100 105 110	
tat ttg ttt tta ttt tga	354

-continued

Tyr Leu Phe Leu Phe
115

<210> SEQ ID NO 52
<211> LENGTH: 117
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic construct
<220> FEATURE:
<221> NAME/KEY: Misc_feature
<222> LOCATION: (1)...(117)
<223> OTHER INFORMATION: OsRHC1 binding partner

<400> SEQUENCE: 52

Met Ala Val Gly Ser Glu Arg Leu Gly Glu Glu Ala Ala Arg Arg Gln
1 5 10 15
Leu Gly Glu Ala Arg Lys Ala Arg Gly Gly Cys Ser Ala Thr Arg Asp
20 25 30
Gly Ala Asp Asp Glu Gly Arg Arg Gln Ile Asn Pro Pro Ser Pro Val
35 40 45
Trp Ser Ser Pro Pro Ser Leu Pro Leu Pro Leu Arg Ser Ala Arg Arg
50 55 60
Gly Thr Gly Gly Gly Arg Arg Pro Pro Phe Pro Leu Ser Ser Gln Ile
65 70 75 80
Arg Pro Val Gly Arg Gly His Arg Arg Gln Arg His Gly Pro Pro Leu
85 90 95
Cys Ser Ser Arg Gly Arg Gln Gly Gly Gly His Arg Ala Val Phe Phe
100 105 110
Tyr Leu Phe Leu Phe
115

The invention claimed is:

1. A recombinant expression system that comprises a nucleotide sequence encoding a protein that has the amino acid sequence of SEQ ID NO:43.

2. A plant or plant cell modified to contain the expression system of claim 1.

3. A method to confer an enhanced ability to resist infections or wounding on a plant, which method comprises modifying said plant to contain the expression system of claim 1.

4. A method to prepare a protein that has the amino acid sequence of SEQ ID NO:43, which method comprises culturing cells that comprise the expression system of claim 1 under conditions wherein said protein is produced and recovering the protein from the culture.

5. A method to confer an enhanced ability to resist infections or wounding on a plant, which method comprises modi-

fyng said plant to contain a recombinant expression system that comprises a nucleotide sequence encoding a protein that has the amino acid sequence of SEQ ID NO:43 or a variant thereof that is at least 95% identical to said amino acid sequence and that confers on plants resistance to infection or wounding, wherein the nucleotide sequence is operatively linked to control systems that effect expression in plant cells, and

wherein said plant is identified as in need of said enhanced ability.

6. The method of claim 5 wherein said variant is at least 98% identical to said amino acid sequence.

7. The method of claim 6 wherein said variant is at least 99% identical to said amino acid sequence.

* * * * *